# Permeation Pathway of Macromolecules and Nanospheres through Skin

Hiroaki Todo,[a] Eriko Kimura,[a] Hirotaka Yasuno,[a] Yoshihiro Tokudome,[a] Fumie Hashimoto,[a] Yoshiaki Ikarashi,[b] and Kenji Sugibayashi*[,a,c]

[a] *Faculty of Pharmaceutical Sciences, Josai University;* [c] *Life Science Research Centre, Josai University; 1–1 Keyakidai, Sakado, Saitama 350–0295, Japan: and* [b] *National Institute of Health Sciences; 1–18–1 Kamiyoga, Setagaya-ku, Tokyo 158–8501, Japan.* Received February 15, 2010; accepted May 31, 2010; published online June 1, 2010

**The permeation pathway of macromolecules and nanospheres through skin was evaluated using fluorescent isothiocyanate (FITC)-dextran (average MW, 4 kDa) (FD-4) and nanospheres (500 nm in diameter) in hairless rat abdominal skin and porcine ear skin as well as a three-dimensional cultured human skin model (cultured skin model). A low molecular hydrophilic compound, sodium fluorescein (FL) (MW, 376 Da), was used for comparison. FL penetrated the stratum corneum and permeated the viable epidermis of hairless rat skin, whereas less permeation of FL was observed through the cultured skin model, suggesting that the primary permeation pathway for the hydrophilic material may be skin appendages through the rat skin. A macromolecular compound, FD-4, was distributed through the hair follicles of the rat skin. In addition, nanospheres were detected in the hair follicles of porcine skin, although no skin permeation was detected. These findings suggest that appendage routes such as hair follicles can be a penetration pathway of macromolecules and nanospheres through skin.**

**Key words** macromolecule; nanosphere; skin permeation pathway; transdermal drug delivery; hair follicle

Delivery of macromolecular compounds and nano-/microparticles has become more realistic due to the recent development of new tools and nanotechnologies for delivery enhancement.[1,2] The administration sites of such macromolecules and nano-/microparticles are supposed to be the mucosa, such as the gastrointestinal (GI) tract,[3] and ophthalmic, nasal and pulmonary[4] mucosa and skin.[5,6] Skin has been paid particular attention as an attractive administration site of these compounds because of its accessibility and easy application. Emulsion droplets, liposomes and other lipophilic carriers[7] containing small molecular active ingredients have already been investigated in the cosmetic field as well as therapeutic drug areas; however, the stratum corneum, the outermost layer of skin, has a primary function to protect the invasion and skin penetration of exogenous substances. Generally, only small molecular compounds less than 500 Da molecular weight are capable of significant passive permeation through the skin barrier (known as the 500 Dalton rule).[8] Thus, skin permeation of macromolecular compounds or nano-/microparticles is very difficult or impossible. Many reports have suggested that large molecules are likely to accumulate on the skin surface or appendages such as hair follicles.[9—12] Nevertheless, few reports have shown a quantitative approach for hair follicular penetration using quantitative skin permeation parameters.

Hairless rat and pig ear skins and three-dimensional cultured human skin model would be useful skin models with and without hair follicles, respectively, to clarify the contribution of hair follicles to skin permeation or the distribution of macromolecules and nanospheres.

In the present study, we selected fluorescent isothiocyanate (FITC)-dextran (average MW, 4 kDa) (FD-4) as a model macromolecular weight compound and 500 nm fluorescent polystyrene latex spheres as model nanospheres, and their potential for skin delivery was investigated by calculating skin permeation parameters or measuring the skin distribution of FD-4 and fluorescent polystyrene latex nanospheres in hairless rat abdominal skin and porcine ear skin as well as a three-dimensional cultured human skin model. Tables 1 and 2 summarize the model penetrant compounds and skin membranes used in this experiment. A low molecular hydrophilic compound, sodium fluorescein (FL) (MW 376 Da), was also used for comparison.

**Theoretical** Skin permeation kinetics is usually evaluated under an assumption that the skin consists of a single barrier membrane against drug permeation; however, generally, the drug-permeable membrane must be classed into three membranes: dissolution–diffusion membrane (Type 1 membrane), porous membrane (Type 2 membrane) and composite membrane (Type 3 membrane) of Type 1 and 2 membranes. Under the assumption that a single barrier of skin is one of these three membranes, the steady state skin permeation rate per unit application area, $dQ/dt$, is expressed using Fick's first law of diffusion as follows:

Type 1 membrane (dissolution–diffusion membrane)

$$\frac{dQ}{dt} = D \cdot \frac{K \cdot C_v}{L} = (KL)\left(\frac{D}{L^2}\right)C_v \tag{1}$$

Type 2 membrane (porous membrane)

$$\frac{dQ}{dt} = \frac{\varepsilon \cdot D_p}{\tau} \cdot \frac{C_v}{L} = (\varepsilon L)\left(\frac{D_p}{\tau L^2}\right)C_v \tag{2}$$

Type 3 membrane (composite membrane)

$$\frac{dQ}{dt} = (1-\varepsilon) \cdot D \cdot \frac{K \cdot C_v}{L} + \frac{\varepsilon \cdot D_p}{\tau} \cdot \frac{C_v}{L} = \left[\{(1-\varepsilon)KL\}\left(\frac{D}{L^2}\right) + (\varepsilon L)\left(\frac{D_p}{\tau L^2}\right)\right]C_v \tag{3}$$

where $C_v$ is the initial concentration of the applied compound, $D$, $K$ and $L$ are diffusion coefficient, partition coefficient and barrier thickness of the membrane, respectively, and $D_p$, $\varepsilon$ and $\tau$ are diffusion coefficients in water-filled pores, average fraction of diffusion area of pores, and tortuosity of the membrane, respectively. In examples 1 and 2, partition parameters and diffusion parameters of the pene-

* To whom correspondence should be addressed. e-mail: sugib@josai.ac.jp

trant are $KL$ and $DL^{-2}$ for the Type 1 membrane and $\varepsilon L$ and $D_p\tau^{-1}L^{-2}$ for the Type 2 membrane, respectively. The permeability coefficient, $P$, can be obtained as a product of the partition parameter and diffusion parameter. Diffusion lag time was obtained by dividing six by the diffusion parameter.

## MATERIALS AND METHODS

**Materials and Animals** Both FL and FD-4 were obtained from Sigma-Aldrich Co., Ltd. (St. Louis, MO, U.S.A.). Fluorescent polystyrene latex nanospheres, Fluoresbrite® yellow green carboxylate microspheres (500 nm in average diameter), were purchased as model nanospheres from Polysciences, Inc. (Warrington, PA, U.S.A.). All other reagents and solvents were of reagent grade or HPLC grade, and used without further purification.

Male hairless rats (WBN/ILA-Ht, *ca.* 200—250 g) were supplied either from Life Science Research Center, Josai University (Sakado, Saitama, Japan) or Ishikawa Experimental Animal Laboratory (Fukaya, Saitama, Japan). Porcine ear skins were from Saitama Experimental Animal Laboratory (Sugito, Saitama, Japan). A three-dimensional cultured human skin model, Living Skin Equivalent-high (LSE-high), was obtained from Toyobo (Osaka, Japan).

**Determination of *n*-Octanol–Water Partition Coefficient** *n*-Octanol–water partition coefficient ($K_{o/w}$) of each fluorescent compound (FL or FD-4) was measured using distilled water–saturated *n*-octanol and *n*-octanol–saturated pH 7.4 phosphate buffered saline (PBS) at 32 °C. *n*-Octanol was added to the same volume of pH 7.4 PBS containing 10 mg/ml of each fluorescent, and the thoroughly mixed solution was equilibrated for 24 h. The aqueous phase was then analyzed using a fluorescence spectrophotometer (RF 5300PC; Shimadzu, Kyoto, Japan) at excitation and emission wavelengths of 490 and 520 nm, respectively. Logarithmic values of the partition coefficients are shown in Table 1.

***In Vitro* Skin Permeation Study** The skin permeation of FL and FD-4 was assessed using excised hairless rat abdominal skin and LSE-high. After the rats had been anesthetized by intraperitoneal injection of sodium pentobarbital (50 mg/kg), the abdominal skin was excised as described in our previous paper.[14] Stripped hairless rat skin was also used after removing the stratum corneum from the abdominal area by stripping 20 times with adhesive tape (Cellophane tape; Nichiban Co., Ltd., Tokyo, Japan). LSE-high was used after removing cultured skin pieces from the plastic insert with a knife. Each skin membrane was mounted in the side-by-side diffusion cell (effective diffusion area: 0.95 $cm^2$),[15,16] and 1.0 mM FL or 0.25 mM FD-4 (2.5 ml each) was applied to the stratum corneum side and the same volume of PBS was applied to the dermal side. Samples of 0.40 ml were taken periodically from the dermal side compartment, and then the same volume of the same solvent was added to keep the volume constant. FL or FD-4 concentration of each sample was determined using a fluorescence spectrophotometer, as explained above. The hairless rat skin and LSE-high surfaces were carefully rinsed with PBS several times to remove FL or FD-4 attached to the stratum corneum 6 h after starting the experiment. The obtained skin sample was embedded in Tissue-Tek® OTC compound (Miles, Inc., Elkhart, IN, U.S.A.) and stored at −80 °C until slicing.

Table 1. Physicochemical Properties of Model Compounds

| Model compounds (abbreviation) | Molecular weight (Da) | Mean particle size or molecular radius | Log $K_{o/w}$ [a] |
|---|---|---|---|
| Sodium fluorescein (FL) | 376 | 0.45 nm[13] (Stokes radius) | −0.615 |
| FITC-dextran (FD-4) | 4000 | 1.4 nm (Stokes radius)[13] | −0.773 |
| Fluorescent polystyrene latex nanospheres (Fluoresbrite) | — | 500 nm | — |

*a*) $K_{o/w}$: *n*-octanol–water partition coefficient.

The skin permeation property of fluorescent polystyrene latex spheres (Fluoresbrite) was evaluated using excised hairless rat skin and excised porcine ear skin, which had been carefully shaved and the underlying excess fatty tissues removed from the dermis. LSE-high was also used to evaluate whether Fluoresbrite permeates the cultured skin. The obtained skin membranes were mounted in a Franz-type diffusion cell[14] (effective diffusion area: 1.77 $cm^2$). Then, 1.0 ml PBS-suspended solution containing Fluoresbrite ($3.64\times10^{10}$ particles/ml for 500 nm spheres) was applied to the stratum corneum surface, whereas 6.0 ml PBS was applied to the dermal side. The skin permeation test was performed at 32 °C over 12 h through hairless rat skin, porcine ear skin and LSE-high, while the receiver solution was continuously stirred with a star-head-type magnetic stirrer. The receiver solution was withdrawn 12 h after beginning the permeation experiment. The skins were then carefully rinsed with PBS several times to remove polystyrene spheres attached to the stratum corneum 12 h after starting the experiment. The obtained skin sample was embedded in Tissue-Tek® OTC compound (Miles, Inc., Elkhart, IN, U.S.A.) and stored at −80 °C until slicing.

All animal experiments were approved by the Institutional Animal Care and Use Committee of Josai University.

**Evaluation of Skin Permeation Kinetics** Steady-state flux was calculated by linear regression of the linear portion of normalized cumulative amount of penetrant permeated *versus* the time-curve (steady state; reached 4—6 h after starting the experiment), and the lag time was calculated from the intercept on the time axis by extrapolation from the steady state skin permeation profile. The normalized cumulative amount of penetrant permeated, $Q_n$, was calculated by dividing the cumulative amount permeated per unit area of skin by the initial concentration of the applied fluorescent compound in the donor compartment.[17] The permeation parameters were obtained by curve fitting the skin permeation data by Scheuplein's equation,[18] which comes from Fick's second law of diffusion. The least squares curve fitting method was performed using Microsoft® Excel Solver.[19] The calculation condition was 100 s for the calculation limit, 100 times for repeated calculation, $10^{-6}$ for accuracy, 5% basic tolerance and $10^{-3}$ for convergence. The pseudo-Newtonian method was used as an algorithm.

**Sectioning of Hairless Rat Skin, Porcine Ear Skin and LSE-High** Hairless rat skin, porcine ear skin and LSE-high embedded in Tissue-Tek® OTC compound were sequentially sliced with a cryostat (CM3050S; Leica, Wetzlar, Germany) to obtain horizontal and vertical 20 $\mu$m-thick sections. The prepared skin sections were observed with a fluorescence mi-

Table 2. Comparison of Skin Thickness and Presence or Absence of Hair Follicles in Several Skin Models

| Skin model | Skin structure constitution | Stratum corneum thickness ($\mu$m) | Epidermis thickness ($\mu$m) | Whole skin thickness (mm) | Skin appendage | Relationship to human skin permeation |
|---|---|---|---|---|---|---|
| Hairless rat skin | Epidermis/dermis | 15.4±3.3[20)] | 23.8±5.3[20)] | 0.86±0.06[20)] | Yes | High |
| Pig skin | Epidermis/dermis | 10.6±0.5 | 52.5±4.1 | 1.2±0.002 | Yes | High |
| LSE-high | Epidermis/dermis | 27.0±0.7 | 31.4±1.3 | 0.12±0.001 | No | High |

croscope (CK40; Olympus Corp., Tokyo, Japan).

**Measurement of Thickness in LSE-High and Porcine Skin** The thicknesses of the stratum corneum, epidermis, and whole skin in LSE-high and porcine skin were microscopically determined from microtomed sections after hematoxylin–eosin staining. Five good sections from each specimen were used to measure the stratum corneum, and whole skin thicknesses were measured by a light micrograph (IX71; Olympus Corp., Tokyo, Japan) and a calibrated ocular micrometer. The thickness of the epidermis was calculated by subtracting the stratum corneum thickness from the whole skin thickness. The thickness of hairless rat skin was cited from our previous paper.[20)]

**Observation of Skin Surface** Shaved hairless rat and porcine ear skins were mounted with adhesive tape on a scanning microscopy (SEM) stage, and the skin surface was observed without coating by a low-vacuum SEM (S-3000N; Hitachi Ltd., Tokyo, Japan).

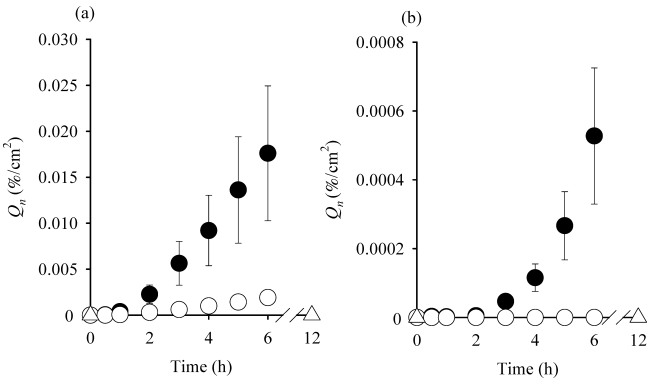


Fig. 1. Cumulative Amount of Hydrophilic Fluorescent Compounds, FL (●), FD-4 (○) and 500 nm Fluoresbrite (△) through Hairless Rat Skin (a) and LSE-High (b)

Normalized cumulative amount of the compounds permeating a percent per unit application area (%/cm$^2$)[17)] was plotted on the vertical axis. Each data point represents the mean±S.E. of 3—4 experiments.

## RESULTS

Many reports have shown that nano-/microspheres could not permeate the healthy stratum corneum.[21)] In our study, therefore, the penetration pathway of hydrophilic fluorescent markers, FL and FD-4, was observed to evaluate the potential penetration of these mal-absorptive materials into skin and the delivery pathway through the skin barrier. The characteristics of model skin membranes (excised hairless rat skin, pig ear skin and LSE-high) are shown in Table 2. The stratum corneum in LSE-high was much thicker than the others. In addition, skin appendages such as sweat ducts and hair follicles could not be observed in LSE-high. Although many structural differences could be found between LSE-high and the others, and the permeation of several compounds (MW 122—236, $-1.5<\log K_{o/w}<2.1$) through LSE-high was about 10 times higher than through hairless rat and human skins, and the permeation rate through LSE-high showed a linear relationship to that through hairless rat and human skins.[14)]

Figure 1a and b show the time course of the normalized cumulative amount of FL and FD-4 that permeated the unit area of excised hairless rat skin and LSE-high, respectively. In these experiments, 1.0 mM FL or 0.25 mM FD-4 (2.5 ml each) was applied to the skin surface to follow skin permeation. Interestingly, both fluorescent markers permeated hairless rat skin, whereas less permeation of FL and no permeation of FD-4 were observed through LSE-high. The $Q_n$ of FL through hairless rat skin was 30-fold higher than through LSE-high.

The typical lag time and subsequent steady state permeation were observed for the permeation of both fluorescents through hairless rat skin. Permeability coefficients of FL and FD-4 were calculated by two methods using steady-state flux (observed value) and the curve fitting method (estimated value). These values are summarized in Table 3. The estimated vales were almost equal to the corresponding observed values and no significant differences appeared in hairless rat data. The calculated values of the permeability coefficient and $Q_n$ of FL through LSE-high were about one-twelfth and one-thirtieth through hairless rat skin, respectively; furthermore, the lag time of FL through LSE-high (estimated value) was about three-fold of that through hairless rat skin. On the other hand, lag times of FL and FD-4 through hairless rat skin were almost the same as those through LSE-high.

Fluoresbrite was also applied to hairless rat skin and LSE-high. Although a Franz-type diffusion cell was used for measuring the skin permeation of Fluoresbrite, no permeation of the nanospheres was detected 12 h after starting the skin permeation experiment (see Fig. 1). No skin permeation was detected when Fluoresbrite was applied to porcine ear skin (data not shown).

Figure 2 shows fluorescent photographs illustrating the skin distribution of FL and FD-4 in hairless rat skin and LSE-high after topical application of these fluorescent markers. High-intensity FL was detected both in the stratum corneum and hair follicles of hairless rat (Fig. 2a), whereas FD-4 was mainly observed in the hair follicles (Fig. 2b). On the other hand, with LSE-high, FL was detected mostly in the stratum corneum and slightly in the viable epidermis (Fig. 2c) and FD-4 was found only on the skin surface (no skin penetration was observed for FD-4) (Fig. 2d). These results also suggest the high contribution of the transfollicular pathway to the transport of mal-absorptive hydrophilic compounds across the skin. In addition, this tendency was more

Table 3. Lag Time and Permeability Coefficients of FL and FD-4 through Excised Hairless Rat Skin or LSE-High

| | | FL | | FD-4 | |
|---|---|---|---|---|---|
| | | Lag time (h) | Permeability coefficient (cm/s) | Lag time (h) | Permeability coefficient (cm/s) |
| Hairless rat | Estimated value[a)] | 2.0±0.16 | $(1.1\pm0.5)\times10^{-8}$ | 2.2±0.01 | $(3.6\pm0.2)\times10^{-9}$ |
| | Observed value[b)] | 1.8±0.20 | $(1.2\pm0.5)\times10^{-8}$ | 2.1±0.01 | $(3.4\pm0.2)\times10^{-9}$ |
| LSE-high | Estimated value[a)] | 5.4±0.4 | $(9.4\pm2.4)\times10^{-10}$ | — | $\ll1.21\times10^{-10}$ [d)] |
| | Observed value[b)] | —[c)] | —[c)] | —[c)] | —[c)] |

*a*) Estimated value was calculated by curve-fitting the time course of the cumulative amount of skin permeation of compounds using Scheuplein's equation.[25)] *b*) Observed value was obtained by the slope of steady-state flux and time–axis intercept of the time course of the cumulative amount of skin permeation of compounds. *c*) No steady state permeation was obtained until 6 h in the skin permeation study. *d*) Estimated value was calculated from lower quantitative limit of FD-4 in receiver cell 6 h after skin permeation study.

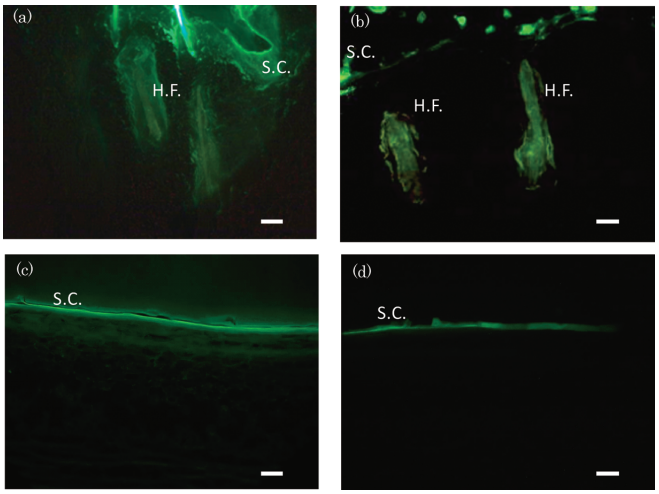


Fig. 2. Histological Observation of Hairless Rat Intact Skin (a, b) and LSE-High (c, d) after Application of FL (a, c) and FD-4 (b, d)

S.C.: Stratum corneum; H.F.: hair follicle. White bars=100 $\mu$m. (a, b): Fluorescence derived from FL or FD-4 was observed on the skin surface of skin and in hair follicles, (c, d): fluorescence derived from FL or FD-4 was observed in shallow areas or only on the surface, respectively, of LSE-high.

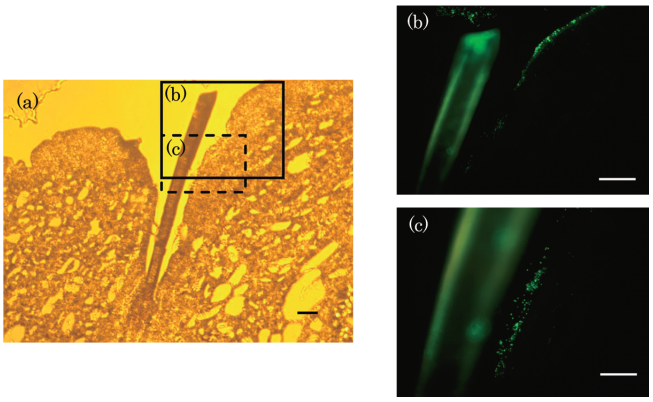


Fig. 3. Histological Observation of Excised Porcine Ear Skin 12 h after Application of 500 nm Fluoresbrite

a: Light micrograph of vertical slice. b and c: Fluorescent micrograph of area b and c in Fig. 3a. Bar=200 $\mu$m. (b, c): Fluoresbrite was observed in infundibulum of the hair follicle and surface of the hair shaft.

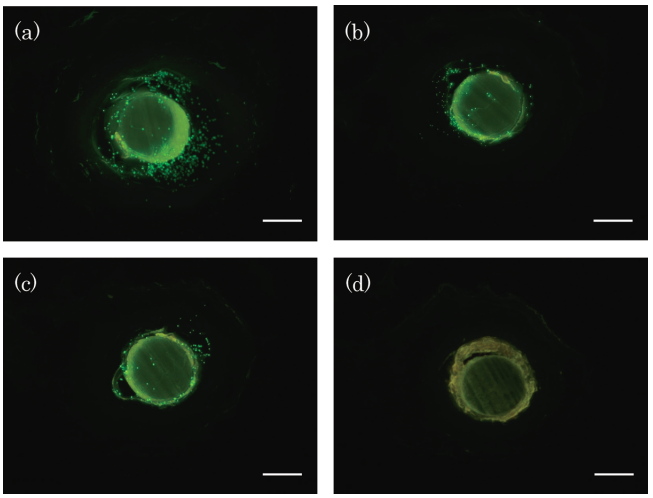


Fig. 4. Localization of 500 nm Fluoresbrite Penetrating the Hair Follicle

Fluorescence images (a—d) are horizontal slices at different depths from the skin surface of excised porcine ear skin 12 h after application of particles. a: *ca.* 20 $\mu$m, b: 40—60 $\mu$m, c: 80—100 $\mu$m, d: 200—220 $\mu$m. Bar=100 $\mu$m. (a): Fluoresbrite was detected on the surface of hair shaft and connective tissue in the follicle, (b, c): number of particles gradually decreased with an increase in depth from the skin surface, (d) only autofluorescence was observed.

marked when using the macromolecular compound. Thus, skin appendages such as hair follicles must be very important for the skin permeation of malabsorptive compounds.

Next, the skin distribution of Fluoresbrite (500 nm in diameter) was investigated after topical application to excised hairless rat skin, excised porcine ear skin and LSE-high. Nanospheres were detected only on the surface of the stratum corneum (data not shown) for hairless rat skin and LSE-high; therefore, a detailed observation was performed using excised porcine skin, since it has much larger hair follicles. Figure 3a shows a light microphotograph of porcine skin (vertical slice of hair follicle area) 12 h after the application of Fluoresbrite, and Figure 3b and c show fluorescent microphotographs of specific parts of the hair follicle area, as explained in Fig. 3a. Many nanospheres were found around the openings of the hair follicle, especially close to the epidermis side and around the hair shaft, as shown in Fig. 3b and c. The penetration depth of Fluoresbrite in the hair follicles was investigated by preparing horizontal slices of the hair follicle area of skin. The thickness of each skin section was adjusted to 20 $\mu$m. Figure 4 shows typical cross-section images of the hair follicle area from the skin surface (0—20 $\mu$m) to dermis side (200—220 $\mu$m) 12 h after application of Fluoresbrite to the excised porcine ear skin. In accordance with the photograph in Fig. 3, nanospheres could be detected in hair follicles, such as the surface of the hair shaft and connective tissue follicles, and the intensity due to nanospheres in the hair follicle gradually decreased with increasing pene-

tration depth; however, nanospheres could not be detected in connective tissue follicles below 200-$\mu$m depth from the skin surface. Only greenish-yellow autofluorescence derived from keratin and melanin was observed in Fig. 4d. This result clearly identified that the nanospheres were distributed or penetrated until about 100-$\mu$m depth, but did not penetrate as far as 200-$\mu$m depth from the skin surface 12 h after application. Thus, macromolecular compounds, such as FD-4 and nanospheres, are probably distributed or penetrate through the transfollicular pathway, although the extent is very marginal.

## DISCUSSION

Three kinds of membranes are frequently utilized to describe the membrane permeation profiles of compounds, as explained in the theoretical section. In the dissolution–diffusion membrane (Type 1 membrane), compounds are dissolved and distributed into the membrane and then diffused in the homogeneous membrane. In the microporous membrane (Type 2 membrane), compounds are diffused across solvent (usually aqueous)-filled pores in the membrane. The third membrane (composite membrane) is the previous two membranes combined.

In the case of hairless rat and porcine skins, the stratum corneum and skin appendage may be the permeation pathway of compounds, especially for low molecular compounds ($\leq$500 Da). Thus, these animal skins would be assumed to be the third membrane. On the other hand, LSE-high would be classified as a dissolution–diffusion membrane, since three-dimensional cultured human skin model has no appendages, such as hair follicles and sweat ducts.

Although LSE-high is such a skin appendage-deficiency model, it was observed in our previous study[14] that logarithmic values of the permeability coefficient, log $P$, of seven drugs through LSE-high were fairly proportional to those through excised hairless rat, pig and human skins, and the partition parameters of LSE-high were almost the same as in other skins. For FL and FD-4 applied to LSE-high, low permeation and no permeation were observed in the present study, while permeation through hairless rat skin was observed. The permeability coefficient, $P$, of FD-4 through LSE-high was calculated from the lower quantitative limit of FD-4 in receiver solution (Table 3). The estimated value was about thirtieth of that of FD-4 through hairless rats. FL (p$K_a$ 1: 4.32, p$K_a$ 2: 6.5)[22] predominantly exists as an ionized form in pH 7.4 PBS, and FD-4 has a high molecular weight; therefore, the $P$-value of FL and FD-4 through LSE-high was much lower than through hairless rat skin.

The $P$-value is a product of the partition parameter ($KL$ or $\varepsilon L$) and diffusion parameter ($DL^{-2}$ or $D_p\tau^{-1}L^{-2}$).[23] To clarify the differences of skin permeation profiles between hairless rat skin and LSE-high, the partition parameter and diffusion parameter were compared.

Table 4 shows the partition and diffusion parameters, which were calculated from curve-fitting the time course of the cumulative amount of FL and FD-4 permeating hairless rat skin and LSE-high. Interestingly, both parameters of LSE-high calculated from FL permeation were not the same as those of hairless rat skin. These differences might reflect the different permeation routes of FL between these skins.

Table 4. Comparison of Partition Parameter and Diffusion Parameter between Hairless Rat Skin and LSE-High

| Compound | Membrane | Partition parameter ($KL$ or $\varepsilon L$) (cm) | Diffusion parameter ($DL^{-2}$ or $D_p\tau^{-1}L^{-2}$) ($s^{-1}$) |
|---|---|---|---|
| FL | Hairless rat | $(4.7\pm1.8)\times10^{-4}$ | $(2.3\pm0.18)\times10^{-5}$ |
| | LSE-high | $(1.1\pm0.31)\times10^{-4}$ | $(8.6\pm0.75)\times10^{-6}$ |
| FD-4 | Hairless rat | $(1.7\pm0.13)\times10^{-4}$ | $(2.0\pm0.06)\times10^{-5}$ |
| | LSE-high | — | — |

$KL$ and $DL^{-2}$ or $\varepsilon L$ and $D_p\tau^{-1}L^{-2}$ were calculated by curve-fitting the skin permeation profile of FL and FD-4 through hairless rat skin and LSE-high.

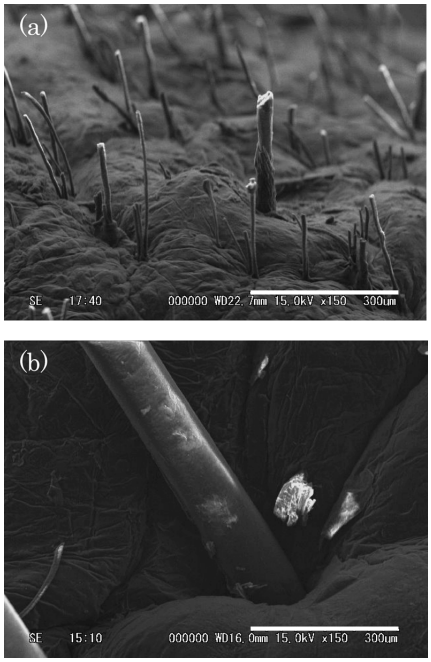


Fig. 5. SEM Observation of Hairless Rat Skin (a) and Porcine Ear Skin (b)

Bar=300 $\mu$m.

Increased partition and diffusion parameters of FL in hairless rat skin might mean that FL was mainly partitioned in pore routes and permeated the pore routes of skin; therefore, skin appendages, such as hair follicles, are the predominant permeation route of FL.

The diffusion parameters of FL and FD-4 in hairless rat skin showed almost the same value due to the slight difference in the cubic root of the molecular weight of these compounds, because the diffusion coefficient is proportional to the reciprocal of the cubic root, which is given by the Stokes–Einstein equation. Therefore, FL and FD-4 must permeate through almost the same permeation pathway of skin. On the other hand, the partition parameter of FD-4 was about one-third that of FL, indicating that FD-4 permeated skin through a more restricted pathway, such as hair follicles, than the FL pathway. In addition to this assumption, the present fluorescence images strongly supported the skin appendage route as a useful pathway for skin permeation and/or the distribution of such macromolecular compounds.

Fluoresbrite did not permeate hairless rat skin or LSE-high. Thus, porcine skin was selected to observe its skin distribution because the pores of hair follicles in porcine skin

are larger than in hairless rat skin (see Fig. 5). Fluoresbrite was especially observed in the infundibulum of hair follicles of porcine skin and no spheres were observed more than 100-$\mu$m depth from the skin surface (Fig. 4). It is reasonable that Fluoresbrite was detected only in the hair follicle because even FD-4, having a smaller molecular radius than nanospheres, was mainly detected in hair follicles.

This indicated that hair follicles are expected to be a useful pathway, not only for macromolecular compounds, but also nanospheres, through the skin barrier. Scheuplein[18] reported that the contribution of the transappendage route to the skin permeation of low molecular compounds must be very low, although the transfollicular pathway would play a very important role in the early stage of skin permeation and distribution. This is because the skin appendage area is only 0.1% of the total skin surface area.[18,24] Further study is necessary to fully elucidate the contribution of hair follicles to the skin permeation or distribution of hydrophilic compounds and nano-/microspheres. This contribution of hair follicles can be assessed using skin permeation parameters, such as $\varepsilon \cdot L$ and $D/\tau \cdot L^2$ as above.

## CONCLUSION

The present study revealed that important role of hair follicles as a permeation pathway or distribution pathway for hydrophilic compounds and nanospheres. Although not only skin features, such as hair density and follicle size, but also physicochemical properties, such as molecular size and *n*-octanol/water partition coefficient of compounds, affect their transfollicular permeation,[25] analysis of the hair follicle contribution to the overall skin permeation of compounds using permeation parameters will help to understand efficient compound targeting of hair follicles.

**Acknowledgement** This study was supported by a Grant-in-Aid for Scientific Research (H20-iyaku-ippan-001) from the Ministry of Health, Labor, and Welfare, Japan.

## REFERENCES

1) Ravi Kumar M. N., *J. Pharm. Pharm. Sci.*, **3**, 234—258 (2000).
2) Bilati U., Allemann E., Doelker E., *Eur. J. Pharm. Biopharm.*, **59**, 375—388 (2005).
3) Takeuchi H., Matsui Y., Sugihara H., Yamamoto H., Kawashima Y., *Int. J. Pharm.*, **303**, 160—170 (2005).
4) Todo H., Iida K., Okamoto H., Danjo K., *J. Pharm. Sci.*, **92**, 2475—2486 (2003).
5) Alvarez-Roman R., Naik A., Kalia Y. N., Guy R. H., Fessi H., *J. Controlled Release*, **99**, 53—62 (2004).
6) Almeida A. J., Souto E., *Adv. Drug Deliv. Rev.*, **59**, 478—490 (2007).
7) Honeywell-Nguyen P. L., Wouter Groenink H. W., Bouwstra J. A., *J. Liposome Res.*, **16**, 273—280 (2006).
8) Bos J. D., Meinardi M. M., *Exp. Dermatol.*, **9**, 165—169 (2000).
9) Lademann J., Weigmann H., Rickmeyer C., Barthelmes H., Schaefer H., Mueller G., Sterry W., *Skin Pharmacol. Appl. Skin Physiol.*, **12**, 247—256 (1999).
10) Toll R., Jacobi U., Richter H., Lademann J., Schaefer H., Blume-Peytavi U., *J. Invest. Dermatol.*, **123**, 168—176 (2004).
11) Trauer S., Patzelt A., Otberg N., Knorr F., Rozycki C., Balizs G., Buttemeyer R., Linscheid M., Liebsch M., Lademann J., *Br. J. Clin. Pharmacol.*, **68**, 181—186 (2009).
12) Teichmann A., Jacobi U., Ossadnik M., Richter H., Koch S., Sterry W., Lademann J., *J. Invest. Dermatol.*, **125**, 264—269 (2005).
13) Mota M. C., Carvalho P., Ramalho J., Leite E., *Int. Ophthalmol.*, **15**, 321—326 (1991).
14) Watanabe T., Hasegawa T., Takahashi H., Ishibashi T., Takayama K., Sugibayashi K., *Altern. Animal Test Experiment.*, **8**, 1—14 (2001).
15) Okumura M., Sugibayashi K., Ogawa K., Morimoto Y., *Chem. Pharm. Bull.*, **37,** 1404—1406 (1989).
16) Obata Y., Takayama K., Maitani Y., Machida Y., Nagai T., *Biol. Pharm. Bull.*, **16**, 312—314 (1993).
17) Sugibayashi K., Hosoya K., Morimoto Y., Higuchi W. I., *J. Pharm. Pharmacol.*, **37**, 578—580 (1985).
18) Scheuplein R. J., *J. Invest. Dermatol.*, **48**, 79—88 (1967).
19) Sugibayashi K., Hayashi T., Matsumoto K., Hasegawa T., *Drug Metab. Pharmacokinet.*, **19**, 352—362 (2004).
20) Sato K., Sugibayashi K., Morimoto Y., *J. Pharm. Sci.*, **80**, 104—107 (1991).
21) Scientific Committee on Consumer Product, "Opinion on Safety on Nanomaterials in Cosmetic Products," 2007.
22) Lauer A. C., Lieb L. M., Ramachandran C., Flynn G. L., Weiner N. D., *Pharm. Res.*, **12**, 179—186 (1995).
23) Okamoto H., Yamashita F., Saito K., Hashida M., *Pharm. Res.*, **6**, 931—937 (1989).
24) Ogiso T., Shiraki T., Okajima K., Tanino T., Iwaki M., Wada T., *J. Drug Target*, **10**, 369—378 (2002).
25) Knorr F., Lademann J., Patzelt A., Sterry W., Blume-Peytavi U., Vogt A., *Eur. J. Pharm. Biopharm.*, **71**, 173—180 (2009).