

Reactions of 26-Iodopseudodiosgenin and 26-Iodopseudodiosgenone with Various Nucleophiles and Pharmacological Activities of the Products

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26-Iodopseudodiosgenin (**8**) and 26-iodopseudodiosgenone (**9**) were reacted with various nucleophiles (KSCN, KOCN, NaCN, NaN₃ and various amines) to give pseudodiosgenin derivatives (**4**, **12**, **16**–**20**, **26**) and pseudodiosgenone derivatives (**5**, **13**, **21**–**25**, **27**), respectively. The reactions of **8** and **9** with KOCN gave the elimination products (**10**) and (**11**), respectively. The reaction of **9** with NaCN gave 5 α ,26- (**14**) and 5 β ,26-dicyanocholestan-3-one (**15**). The reaction of **8** with NaN₃ gave triazepine derivative (**30**), while that of **9** gave 26-azido-pseudodiosgenone (**31**). Compound **31** was converted into triazepine derivative (**32**) by heating at 120 °C. The cytotoxicity of the pseudodiosgenins and pseudodiosgenones on P-gp-underexpressing HCT 116 cells and P-gp-overexpressing Hep G2 cells was examined by MTT assay. Pseudodiosgenins **2**, **4**, **12** and **30** showed strong cytotoxic activity (IC₅₀ values: 2.6 \pm 0.3–6.7 \pm 1.4 μ M), as did pseudodiosgenones **3**, **5**, **11**, **13**, **21**–**25** and **27** (IC₅₀ values: 1.3 \pm 0.3–6.4 \pm 0.3 μ M) toward HCT 116 cells. Pseudodiosgenins **12**, **16** and **30** (IC₅₀ values: 1.2 \pm 0.7–2.2 \pm 0.6 μ M) and pseudodiosgenones **22**, **23**, **25** and **27** (IC₅₀ values: 0.6 \pm 0.1–2.5 \pm 0.3 μ M) were highly cytotoxic to Hep G2 cells. Compounds **3** and **27** showed efficient antibacterial activity (MIC: 15.6, 10.4 μ g/ml) and (MIC: 7.8, 15.6 μ g/ml) against *Bacillus subtilis* and *Staphylococcus aureus*, respectively.

Key words pseudodiosgenin; pseudodiosgenone; cytotoxic activity; P-glycoprotein; antibacterial activity

Saponins have various pharmacological effects such as anti-tumor,¹⁾ anti-allergic,²⁾ anti-inflammatory,³⁾ and anti-HIV^{4,5)} activity. Previously, we reported the structure–activity relationships for the anti-hepatitis^{6–12)} and anti-HIV activity¹³⁾ of various glycosides constructed using a combination of different sugars and aglycons synthetically derived from glycyrrhetic acid.¹⁴⁾ Furthermore, we compared the cytotoxicity of various synthetic spirostanols and heterospirostanols¹⁵⁾ derived from diosgenin (**1**). In the investigation of cytotoxicity, we found that 26-cyanoselenopseudodiosgenone (**3**) (an opened F ring structure) showed marked activity. This finding led us to attempt to prepare furostane derivatives having various substituents containing other hetero atoms such as sulfur and nitrogen at position 26 and to evaluate the pharmacological activity. The present study deals with the reactions of 26-iodopseudodiosgenin (**8**) and 26-iodopseudodiosgenone (**9**) with various nucleophiles to obtain various pseudodiosgenin and pseudodiosgenone derivatives, and evaluations of cytotoxicity of the derivatives using cancerous HCT 116 and Hep G2 cells. Furthermore, antibacterial activity against *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*, and antifungal activity against *Candida albicans* and *Aspergillus niger* are investigated.

Results and Discussion

We already reported the synthesis of 26-cyanoselenopseudodiosgenin (**2**) and 26-cyanoselenopseudodiosgenone (**3**)¹⁵⁾ according to the methods reported by Uhle^{16–18)} with minor modifications. This time, the pseudodiosgenin analogs (**4**) and (**6**) and pseudodiosgenone analogs (**5**) and (**7**) having a SCN and OCN group, respectively, and pseudodiosgenin and pseudodiosgenone derivatives having various amino substituents at position 26 were prepared. For the synthesis, 26-iodopseudodiosgenin **8** and 26-iodopseudodiosgenone **9**¹⁵⁾ were used as starting materials. The reaction conditions and

products of the reactions of **8** and **9** with various nucleophiles (runs 1–21) are listed in Table 1.

The reaction of **8** with potassium thiocyanate (KSCN) (run

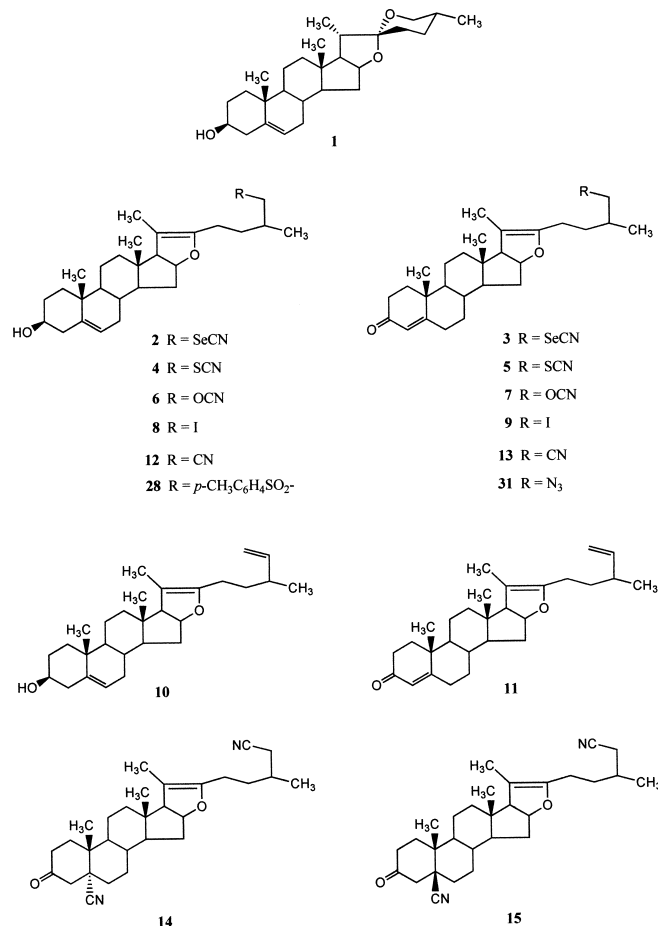


Fig. 1. Structures of Compounds **1**–**15**, **28** and **31**

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1) gave 26-cyanothiopseudodiosgenin **4** in 40% yield. The reaction of **9** with KSCN (run 11) gave 26-cyanothiopseudodiosgenone **5** in 79% yield.

To prepare compounds **6** and **7**, each of the iodides **8** and **9** was reacted with potassium cyanate (KOCN) (runs 2, 12, respectively). However, rather than **6** and **7**, the reaction gave the elimination products **10** and **11** in yields of 25 and 40%. These results may be due to the weaker nucleophilicity and the stronger basicity of the ^-OCN group than ^-SCN and ^-SeCN groups.

26-Cyanopseudodiosgenin (**12**) and 26-cyanopseudo-

diosgenone (**13**), which have no S and Se atoms, were synthesized for comparison of their pharmacological activities with those of **2**–**5**. The reaction of iodide **8** with sodium cyanide (NaCN) at 60 °C for 16 h (run 3) gave compound **12** in 75% yield. On the other hand, the reaction of iodide **9** with NaCN at 60 °C for 12 h (run 13) gave dicyanides **14** (18% yield) and **15** (18% yield) and unisolated products. Thus, as the reaction of **9** with NaCN at 60 °C for 12 h gave dicyanides but not monocyanides such as **13**, the reaction was run for less time (2.5 h) at the same temperature (run 14) to give **13**, **14** and **15** in yields of 33, 12 and 17%, respectively.

The introduction of various amino groups at position 26 of pseudodiosgenins and pseudodiosgonones was performed by reacting iodides **8** and **9** with several kinds of amines such as diethyl amine, pyrrolidine, piperidine, morpholine, 1-methylpiperazine and imidazole as well as sodium azide. All reactions were carried out at 60 °C. The reactions of **8** with diethyl amine, pyrrolidine, piperidine, morpholine and 1-methylpiperazine (runs 4–8, respectively) gave the corresponding amines **16**–**20** in yields of 58, 70, 80, 54 and 62%, respectively. The reactions of **9** with diethyl amine, pyrrolidine, piperidine, morpholine and 1-methylpiperazine (runs 15–19, respectively) gave compounds (**21**–**25**) in 64, 27, 79, 73 and 61% yields, respectively. The reaction of **8** with imidazole in the presence of K_2CO_3 (run 9) gave (**26**) and **10** in 33 and 47% yields, respectively, and that of **9** with imidazole in the presence of K_2CO_3 (run 20) gave (**27**) and **11** in 18 and 27% yields, respectively.

Uhle¹⁸⁾ reported that 26-*p*-tosylpseudodiosgenin (**28**) reacted with sodium azide to give 3 β -hydroxy-22(27)-imino-20(N)-azo-25 α -5-furostene (**29**). To obtain **29**, iodide **8** was used as a starting material instead of **28**; compound **8** was reacted with sodium azide at 60 °C for 12 h (run 10) according to the method reported by Uhle to give 22-((6*R*)-4,5,6,7-tetrahydro-6-methyl-1*H*-1,2,3-triazepine-4-yl)-pseudodiosgenin (**30**) in 43% yield, which was a different product from **29**. The 1H -NMR spectrum of **30** exhibited only one proton

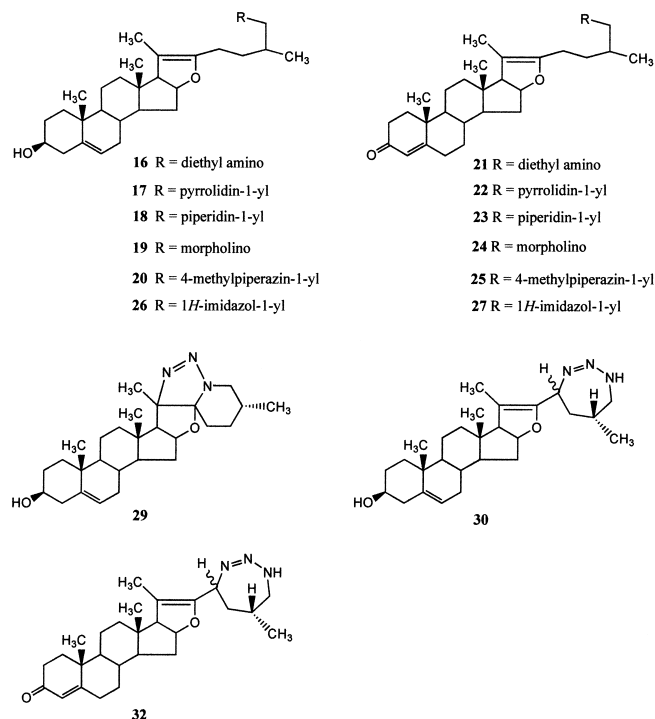


Fig. 2. Structures of Compounds **16**–**27**, **29**, **30** and **32**

Table 1. The Reaction Conditions and Products of the Substitutions of Compounds **8** and **9** with Various Nucleophiles^{a)}

| Run | Substrate | Nucleophile (eq) | Reaction time (h) | Product(s) (%) |
|-----|-----------|-----------------------------|-------------------|--|
| 1 | 8 | KSCN (4) | 12 | 4 (40) |
| 2 | 8 | KOCN (4) | 12 | 10 (25) |
| 3 | 8 | NaCN (4) | 16 | 12 (75) |
| 4 | 8 | Diethyl amine (4) | 12 | 16 (58) |
| 5 | 8 | Pyrrolidine (4) | 1 | 17 (70) |
| 6 | 8 | Piperidine (4) | 2 | 18 (80) |
| 7 | 8 | Morpholine (4) | 5 | 19 (54) |
| 8 | 8 | 1-Methylpiperazine (4) | 4 | 20 (62) |
| 9 | 8 | Imidazole (4) ^{b)} | 3 | 26 (33), 10 (47) |
| 10 | 8 | NaN ₃ (4) | 12 | 30 (43) |
| 11 | 9 | KSCN (4) | 12 | 5 (79) |
| 12 | 9 | KOCN (2) | 24 | 11 (40) |
| 13 | 9 | NaCN (4) | 12 | 14 (18), 15 (18) |
| 14 | 9 | NaCN (4) | 2.5 | 13 (33), 14 (12), 15 (17) |
| 15 | 9 | Diethyl amine (4) | 3 | 21 (64) |
| 16 | 9 | Pyrrolidine (4) | 2 | 22 (27) |
| 17 | 9 | Piperidine (4) | 12 | 23 (79) |
| 18 | 9 | Morpholine (4) | 12 | 24 (73) |
| 19 | 9 | 1-Methylpiperazine (4) | 12 | 25 (61) |
| 20 | 9 | Imidazole (4) ^{b)} | 3 | 27 (18), 11 (27) |
| 21 | 9 | NaN ₃ (4) | 12 | 31 (61) |

a) Reaction of **9** with KOCN was performed at 30 °C (run 12) and the other reactions at 60 °C. b) Potassium carbonate (10 eq) was added in these reactions.

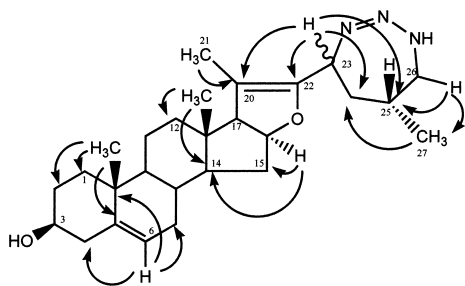


Fig. 3. ^1H – ^{13}C Long-Range Correlations Observed for Compound **30**

signal due to H-23 at δ 4.31 (1H, t, $J=7.6$ Hz), which suggests that the methylene group (CH_2) at position 23 in **8** changes into a methyne group (CH) in **30**. Two vinyl carbon signals due to C-5 and C-6 were observed at δ 140.8 and 121.2, respectively, and two vinyl carbon signals due to C-20 and C-22 of the enol moiety in the E ring were observed at δ 77.6 and 162.9 in the ^{13}C -NMR spectrum, respectively. In the heteronuclear multiple bond connection (HMBC) spectrum of **30** (Fig. 3), correlations between the H-23 signal at δ 4.31 and carbon signals at δ 77.6, 162.9, 29.5 and 34.2 due to C-20, C-22, C-24 and C-25, respectively, were observed. The configuration at position 23 was not determined in this study. The same reaction of iodide **9** with sodium azide as **8** under the same reaction conditions (run 19) gave only the azido product (**31**). Compound **31** was heated at 120 °C for 20 h to give 22-((6*R*)-4,5,6,7-tetrahydro-6-methyl-1*H*-1,2,3-triazepin-4-yl)-pseudodiosgenone (**32**) in 31% yield. These results suggest that products **30** and **32** are obtained *via* 26-azido derivatives by the cyclization of the azido group.

The cytotoxicity of synthetic pseudodiosgenin derivatives **2**,¹⁵⁾ **4**, **10**, **12**, **16**–**20**, **26** and **30**, pseudodiosgenone derivatives **3**,¹⁵⁾ **5**, **11**, **13**, **21**–**25** and **27**, and 5,6-dicyanocholestan-3-one derivatives **14** and **15** was evaluated using human colorectal (HCT 116) and human hepatoma (Hep G2) cancer cell lines. The former cells express very little MDR 1 (P-glycoprotein; P-gp),¹⁹⁾ while the latter cells overexpress P-gp.²⁰⁾ P-gp acts as an efflux pump to remove several antitumor agents, Ca^{2+} antagonists, cyclosporine, digoxin and other compounds from cells.²¹⁾ The cytotoxic activity of each pseudodiosgenin and pseudodiosgenone derivative was estimated by MTT assay,²²⁾ and IC_{50} values calculated based on the percent inhibition of growth are listed in Tables 2 and 3, respectively.

First, the effects on HCT 116 cells of pseudodiosgenins **2**, **4**, **10**, **12**, **16**–**20** and **26** (Table 2) were compared with those of pseudodiosgenones **3**, **5**, **11**, **13**, **21**–**25** and **27** (Table 3) which had the same side chain structures as the corresponding pseudodiosgenins. Among the pseudodiosgenins, only **2**, **4** and **12** showed strong cytotoxic activity (IC_{50} values: 2.6 ± 0.3 – 6.7 ± 1.4 μM). In contrast, the pseudodiosgenones were highly cytotoxic (IC_{50} values: 1.3 ± 0.3 – 6.4 ± 0.3 μM). Thus, the presence of an α,β -unsaturated ketone group on the A ring in steroidal compounds may contribute to the cytotoxic activity.

Previously we reported that a comparison of spirostanol derivatives with furostanol derivatives revealed the presence of a cyanoseleno group at position 26 of the furostanol derivative to be important for strong cytotoxic activity.¹⁵⁾ The cytotoxicity in HCT 116 cells of 26-cyanohetero compounds

Table 2. Cytotoxic Activities of Pseudodiosgenin Derivatives **2**, **4**, **10**, **12**, **16**–**20**, **26** and **30**

| Compound | $\text{IC}_{50} \pm \text{S.D. } (\mu\text{M})^a)$ | |
|-----------|--|------------------|
| | HCT 116 cells | Hep G2 cells |
| 2 | 2.6 ± 0.3 | 4.5 ± 1.5 |
| 4 | 5.3 ± 0.7 | 53.5 ± 5.5 |
| 10 | 14.8 ± 0.7 | 39.8 ± 7.4 |
| 12 | 6.7 ± 1.4 | 1.2 ± 0.7 |
| 16 | 11.6 ± 1.3 | 2.2 ± 0.6 |
| 17 | 49.9 ± 6.8 | 34.7 ± 6.9 |
| 18 | 13.5 ± 0.6 | 6.4 ± 2.8 |
| 19 | 106.9 ± 5.6 | 133.1 ± 27.0 |
| 20 | 32.3 ± 5.1 | 16.3 ± 4.1 |
| 26 | 16.6 ± 1.7 | 8.7 ± 2.8 |
| 30 | 5.4 ± 0.9 | 2.1 ± 0.5 |

a) IC_{50} values are the concentrations at which 50% of the cells are inhibited from growing. S.D., standard deviation. Each experiment was performed in duplicate wells, and drug treatments were performed separately three times.

Table 3. Cytotoxic Activities of Pseudodiosgenone Derivatives **3**, **5**, **11**, **13**, **21**–**25** and **27** and 5,6-Dicyanocholestan-3-one Derivatives **14** and **15**

| Compound | $\text{IC}_{50} \pm \text{S.D. } (\mu\text{M})^a)$ | |
|-----------|--|----------------|
| | HCT 116 cells | Hep G2 cells |
| 3 | 1.7 ± 0.4 | 4.9 ± 1.2 |
| 5 | 1.3 ± 0.3 | 3.5 ± 0.8 |
| 11 | 3.7 ± 0.5 | 6.0 ± 0.9 |
| 13 | 6.4 ± 0.3 | 5.5 ± 1.4 |
| 21 | 3.7 ± 0.5 | 2.9 ± 0.8 |
| 22 | 2.8 ± 0.2 | 2.4 ± 0.6 |
| 23 | 1.9 ± 0.3 | 0.6 ± 0.1 |
| 24 | 3.2 ± 0.3 | 3.7 ± 1.9 |
| 25 | 2.6 ± 0.3 | 1.6 ± 0.5 |
| 27 | 2.6 ± 0.4 | 2.5 ± 0.3 |
| 14 | 5.9 ± 0.7 | 12.4 ± 4.8 |
| 15 | 38.6 ± 6.1 | 30.4 ± 6.1 |

a) IC_{50} values are the concentrations at which 50% of the cells are inhibited from growing. S.D., standard deviation. Each experiment was performed in duplicate wells, and drug treatments were performed separately three times.

was compared with that of 26-cyano compounds. 26-Cyanoseleno- **2** and 26-cyanothiopseudodiosgenin **4** were more potent (IC_{50} values: 2.6 ± 0.3 , 5.3 ± 0.7 μM , respectively) than 26-cyanopseudodiosgenin **12** (IC_{50} value 6.7 ± 1.4 μM). Similarly, 26-cyanoseleno- **3** and 26-cyanothiopseudodiosgenone **5** were more potent (IC_{50} values: 1.7 ± 0.4 , 1.3 ± 0.3 μM , respectively) than 26-cyanopseudodiosgenone **13** (IC_{50} value: 6.4 ± 0.3 μM). These results suggested that the presence of hetero atoms such as selenium and sulfur in the side chain of steroidal compounds enhances the cytotoxicity. However, 25-enepseudodiosgenone **11** showed strong activity (IC_{50} value: 3.7 ± 0.5 μM), though 25-enepseudodiosgenin **10** was less active (IC_{50} value: 14.8 ± 0.7 μM) than **2**–**5**. Therefore, it seems that the presence of substituents such as SeCN, SCN and CN is not essential for enhancement of the cytotoxic activity of steroidal compounds. 5 α ,26-Dicyanocholestan-3-one **14** was more potent (IC_{50} value: 5.9 ± 0.7 μM) than 5 β ,26-dicyanocholestan-3-one **15** (38.6 ± 6.1 μM) in HCT 116 cells.

Next, the cytotoxicity on Hep G2 cells of pseudodiosgenins **2**, **4**, **10**, **12**, **16**–**20** and **26** (Table 2) was compared with that of pseudodiosgenones **3**, **5**, **11**, **13**, **21**–**25** and **27**

(Table 3). In spite of the presence of an efflux system such as P-gp, some compounds still showed strong cytotoxic activity. In the case of the pseudodiosgenins, although the effect of **2** (IC_{50} value: $4.5 \pm 1.5 \mu M$) and **4** (IC_{50} value: $53.5 \pm 5.5 \mu M$) on Hep G2 cells was weaker than that (IC_{50} values: $2.6 \pm 0.3 \mu M$ for **2** and $5.3 \pm 0.7 \mu M$ for **4**) on HCT 116 cells, **2** still exhibited strong activity toward Hep G2 cells. On the other hand, **23** and **30** had more of an effect on Hep G2 cells (IC_{50} values: 0.6 ± 0.1 , $2.1 \pm 0.5 \mu M$, respectively) than HCT 116 cells (1.9 ± 0.3 , $5.4 \pm 0.9 \mu M$, respectively). Furthermore, pseudodiosgenins **12**, **16** and **18** (IC_{50} values: 1.2 ± 0.7 , 2.2 ± 0.6 , $6.4 \pm 2.8 \mu M$, respectively) and pseudodiosgenones **22**, **25** and **27** (1.6 ± 0.5 — $2.5 \pm 0.3 \mu M$) had strong effects even on the P-gp-overexpressing Hep G2 cells. Notably, compound **23** exhibited marked cytotoxic activity (IC_{50} values: $0.6 \pm 0.1 \mu M$) (Tables 2, 3).

Previously we reported¹⁵ that 26-cyanoselenopseudodiosgenone **3** exhibits efficient antibacterial activity against *B. subtilis* 168 UV as measured by an image analysis of cell motion.^{23,24} In this study, antibacterial and antifungal activities of pseudodiosgenins **2**, **4**, **10**, **12**, **16**—**20**, **26** and **30**, pseudodiosgenones **3**, **5**, **11**, **13**, **21**—**25** and **27**, and two 5,26-dicyanocholestan-3-ones **14** and **15**, were investigated according to the protocols of the national committee for clinical laboratory standards (NCCLS).^{25–27} Among these compounds, pseudodiosgenones **3** and **27** were effective against gram-positive *B. subtilis* and *S. aureus* (MIC: 7.8—15.6 $\mu g/ml$) (Table 4). Pseudodiosgenones **22** and **25**, and pseudodiosgenin **16** were also active against both gram-positive bacteria, although their MIC (31.3—62.5 $\mu g/ml$) values were higher than those of the former two compounds. On the other hand, none of the compounds tested revealed any antibacterial activity against gram-negative bacteria (*E. coli* and *P. aeruginosa*). The pseudodiosgenin **16** was the only compound which showed antifungal activity against *C. albicans* (MIC: 62.5 $\mu g/ml$) and *A. niger* (62.5 $\mu g/ml$), though its effects were weak. The other pseudodiosgenins, pseudodiosgenones and 5,26-dicyanocholestan-3-ones used in this study showed no effective antibacterial and antifungal activities; their values were higher than 100 $\mu g/ml$.

Conclusions

Reactions of 26-iodopseudodiosgenin **8** and 26-iodopseudodiosgenone **9** with various nucleophiles were performed.

The cytotoxicity of the products were evaluated using HCT 116 and Hep G2 cancer cell lines. Although only four pseudodiosgenins, **2**, **4**, **12** and **30**, had strong cytotoxic effects (IC_{50} values: 2.6 ± 0.3 — $6.7 \pm 1.4 \mu M$) on HCT 116 cells, all of the pseudodiosgenones were highly active (IC_{50} values: 1.3 ± 0.3 — $6.4 \pm 0.3 \mu M$) against the same cells. These results suggest that the presence of an α,β -unsaturated ketone group on the A ring in steroidal compounds contributes to the cytotoxic effect on HCT 116 cells. Against Hep G2 cells overexpress P-gp, pseudodiosgenins **12**, **16** and **30** (IC_{50} values of 1.2 ± 0.7 , 2.2 ± 0.6 , $2.1 \pm 0.5 \mu M$, respectively) and pseudodiosgenones **22**, **23**, **25** and **27** (IC_{50} values of 2.4 ± 0.6 , 0.6 ± 0.1 , 1.6 ± 0.5 , $2.5 \pm 0.3 \mu M$, respectively) showed potent activities. Thus, these compounds were little susceptible to an efflux system such as P-gp in Hep G2 cells. Of all the compounds investigated, **23**, 26-(piperidine-1-yl)pseudodiosgenone, had the most potent cytotoxic effect on P-gp-

Table 4. Antibacterial and Antifungal Activities (MIC, $\mu g/ml$) of Compounds **3**, **16**, **22**, **25** and **27**^{a)}

| Compound | <i>B. subtilis</i> | <i>S. aureus</i> | <i>C. albicans</i> | <i>A. niger</i> |
|-----------|--------------------|------------------|--------------------|-----------------|
| 3 | 15.6 | 10.4 | >100 ^{b)} | >100 |
| 16 | 31.3 | 62.5 | 62.5 | 62.5 |
| 22 | 62.5 | 41.7 | >100 | >100 |
| 25 | 31.3 | 31.3 | >100 | >100 |
| 27 | 7.8 | 15.6 | >100 | >100 |

a) Each experiment was performed in duplicate wells. Drug treatments were performed separately three times. MIC values greater than 100 $\mu g/ml$ are indicated as >100.

overexpressing Hep G2 cells, and also showed marked activity on HCT 116 cells (IC_{50} value: $1.9 \pm 0.3 \mu M$).

The antibacterial and antifungal activity also were investigated. Compounds **3** and **27** were effective against the gram-positive bacteria tested, while the other compounds showed little or no activity against these bacteria. Activity against gram-negative bacteria was not observed for all compounds tested. Only compound **16** showed weak antifungal activity toward *C. albicans* and *A. niger*. The other compounds showed no antifungal activity.

Experimental

General Methods The chemicals and solvents were of reagent grade and obtained from commercially sources. Melting points (mp) were determined using a Yanagimoto micromelting point apparatus and were uncorrected. Kieselgel 60 F₂₅₄ (Merck) was utilized for the thin-layer chromatography (TLC). Spots were detected by spraying 1:9 Ce(SO₄)₂–10% H₂SO₄ reagent, and heating the plate at 250 °C for 3 min. Column chromatographies were carried out using a Silica gel 60 (Merck). Then, the eluates were monitored using TLC. An SSC-6300 HPLC instrument (Senshu Scientific Co. Ltd.) was employed for analytical HPLC (DOCOSIL, 10×250 mm; flow rate, 1.0 ml min^{−1}; column temperature 40 °C). The SSC-6300 was further equipped with an SSC autinjector 6310 and an SSC fraction collector 6320 for preparative HPLC (DOCOSIL or DOCOSIL-B, 10×250 mm; flow rate, 1.0 ml min^{−1}; column temperature 40 °C). ¹H- and ¹³C-NMR spectra at 500 and 125 MHz, respectively, as well as ¹H–¹H and ¹H–¹³C COSY, DEPT and HMBC spectra were obtained with a JEOL JNM-A500 FT-NMR spectrometer. Tetramethylsilane was used as an internal standard. Chemical shifts were given in ppm. Multiplicities of ¹H-NMR signals were indicated as s (singlet), d (doublet), dd (doublet of doublet), t (triplet) and m (multiplet). FAB-MS and high resolution mass (HR-MS) spectra were recorded on a JEOL LMS-DX 300 mass spectrometer.

General Procedure for Preparing 26-Substituted Pseudodiosgenins and 26-Substituted Pseudodiosgenones Reactions of compounds **8** and **9** with various nucleophiles were performed under the conditions listed in Table 1. To a solution of **8**¹⁵ in DMF (15 ml), was added nucleophile and the reaction mixture was stirred. The reaction mixture was poured into ice-cold water (50 ml) and extracted with dichloromethane (30 ml×4). The extracts were washed with brine, dried over anhydrous sodium sulfate and then filtered. The filtrate was evaporated to give a residue.

26-Cyanothiopseudodiosgenin (4) The reaction of **8** (200 mg, 0.38 mmol) with KSCN (148 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The resulting residue was purified by preparative HPLC (15% H₂O–acetone) to give **4** (70 mg; 40% yield) as a semi-solid. FAB-MS: *m/z* 478 [M+Na]⁺; FAB-MS *m/z*: 455.2853 (Calcd for C₂₈H₄₁O₂NS: 455.2858); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.34 (1H, d, *J*=5.5 Hz, H-6), 4.75 (1H, ddd, *J*=10.1, 7.9, 5.8 Hz, H-16), 3.51 (1H, m, H-3), 3.03 (1H, dd, *J*=12.8, 5.5 Hz, H-26a), 2.80 (1H, dd, *J*=12.8, 7.6 Hz, H-26b), 2.48 (1H, d, *J*=10.1 Hz, H-17), 1.60 (3H, s, 21-CH₃), 1.07 (3H, d, *J*=6.4 Hz, 27-CH₃), 1.02 (3H, s, 19-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 150.5 (C-22), 140.8 (C-5), 121.2 (C-6), 112.7 (SCN), 104.4 (C-20), 84.3 (C-16), 71.5 (C-3), 64.0 (C-17), 54.9 (C-14), 50.0 (C-9), 43.2 (C-13), 42.1 (C-4), 41.1 (C-26), 39.4 (C-1), 37.2 (C-12), 36.5 (C-10), 34.0 (C-15), 33.2 (C-25), 32.5 (C-24), 32.1 (C-7), 31.5 (C-2), 31.1 (C-8), 23.0 (C-11), 20.9 (C-23), 19.3 (C-19), 18.3 (C-27), 13.9 (C-18), 11.6 (C-21).

Pseudodiosgenin-25(26)-ene (10) The reaction of **8** (200 mg, 0.38 mmol) with KOCN (123 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The resulting residue was purified by preparative HPLC (15% H₂O–acetone) to give **10** (38 mg; 25% yield) as a powder, mp 86–88 °C. FAB-MS: *m/z* 419 [M+Na]⁺; FAB-MS *m/z*: 396.3022 (Calcd for C₂₇H₄₀O₂: 396.3029); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.35 (1H, dt, *J*=5.5, 2.1 Hz, H-6), 4.74 (1H, ddd, *J*=10.1, 7.6, 5.8 Hz, H-16), 4.70 (2H, m, H-26), 3.52 (1H, m, H-3), 2.48 (1H, d, *J*=10.4 Hz, H-17), 1.73 (3H, s, 27-CH₃), 1.59 (3H, s, 21-CH₃), 1.03 (3H, s, 19-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 151.3 (C-22), 145.4 (C-25), 140.8 (C-5), 121.4 (C-6), 109.9 (C-26), 103.7 (C-20), 84.3 (C-16), 71.7 (C-3), 64.2 (C-17), 55.0 (C-14), 50.1 (C-9), 43.2 (C-13), 42.2 (C-4), 39.5 (C-1), 37.2 (C-12), 36.6 (C-10), 35.2 (C-24), 34.1 (C-15), 32.2 (C-7), 31.6 (C-2), 31.2 (C-8), 24.4 (C-11), 22.4 (C-27), 21.0 (C-23), 19.4 (C-19), 13.9 (C-18), 11.6 (C-21).

26-Cyanopseudodiosgenin (12) The reaction of **8** (200 mg, 0.32 mmol) with NaCN (75 mg, 1.53 mmol) was performed using standard procedures to obtain a residue. The resulting residue was subjected to preparative HPLC (15% H₂O–acetone) to give **12** (121 mg; 75% yield) as a powder, mp 132 °C. FAB-MS: *m/z* 446 [M+Na]⁺; FAB-MS *m/z*: 423.3137 (Calcd for C₂₈H₄₁O₂N: 423.3137); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.34 (1H, d, *J*=5.2 Hz, H-6), 4.73 (1H, ddd, *J*=10.1, 7.9, 5.8 Hz, H-16), 3.51 (1H, m, H-3), 2.48 (1H, d, *J*=10.1 Hz, H-17), 2.35 (1H, dd, *J*=16.8, 5.2 Hz, H-26a), 2.24 (1H, dd, *J*=16.7, 7.3 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.08 (3H, d, *J*=6.7 Hz, 27-CH₃), 1.02 (3H, s, 19-CH₃), 0.68 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 150.5 (C-22), 140.7 (C-5), 121.2 (C-6), 118.7 (CN), 104.2 (C-20), 84.3 (C-16), 71.4 (C-3), 64.0 (C-17), 54.9 (C-14), 49.9 (C-9), 43.1 (C-13), 42.1 (C-4), 39.4 (C-1), 37.1 (C-12), 36.5 (C-10), 34.0 (C-15), 33.1 (C-24), 32.0 (C-7), 31.4 (C-2), 31.0 (C-8), 29.8 (C-25), 24.2 (C-26), 23.0 (C-11), 20.9 (C-23), 19.3 (C-19), 19.1 (C-27), 13.8 (C-18), 11.5 (C-21).

26-Diethylaminopseudodiosgenin (16) The reaction of **8** (200 mg, 0.38 mmol) with diethyl amine (112 mg, 1.53 mmol) was performed using standard procedures to obtain a residue. The resulting residue was subjected to column chromatography (a gradient of 0–90% acetone in toluene) to give **16** (104 mg; 58% yield) as a powder, mp 115–117 °C. FAB-MS: *m/z* 492 [M+Na]⁺; FAB-MS *m/z*: 470.3992 (Calcd for C₃₁H₅₁O₂N+H: 470.3998); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.34 (1H, d, *J*=5.2 Hz, H-6), 4.72 (1H, ddd, *J*=10.1, 7.9, 5.8 Hz, H-16), 3.51 (1H, m, H-3), 2.47 (4H, CH₂×2, q, *J*=7.0 Hz, NCH₂CH₃), 2.46 (1H, d, *J*=10.4 Hz, H-17), 2.23 (1H, dd, *J*=12.5, 5.8 Hz, H-26a), 2.09 (1H, dd, *J*=12.5, 7.6 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.02 (3H, s, 19-CH₃), 0.98 (6H, CH₃×2, t, *J*=7.0 Hz, NCH₂CH₃), 0.90 (3H, d, *J*=6.4 Hz, 27-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 152.0 (C-22), 140.8 (C-5), 121.2 (C-6), 103.2 (C-20), 84.1 (C-16), 71.4 (C-3), 64.1 (C-17), 60.1 (C-26), 55.0 (C-14), 50.0 (C-9), 47.4 (NCH₂CH₃), 43.2 (C-13), 42.2 (C-4), 39.5 (C-1), 37.2 (C-12), 36.5 (C-10), 34.1 (C-15), 32.7 (C-24), 32.1 (C-7), 31.5 (C-2), 31.2 (C-8), 31.1 (C-25), 23.4 (C-11), 20.9 (C-23), 19.3 (C-19), 18.3 (C-27), 13.8 (C-18), 11.6 (C-21) and NCH₂CH₃).

26-(Pyrrolidin-1-yl)pseudodiosgenin (17) The reaction of **8** (200 mg, 0.38 mmol) with pyrrolidine (108 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The recrystallization from acetone gave **17** (125 mg; 70% yield) as needles, mp 181–183 °C. FAB-MS: *m/z* 468 [M+H]⁺; FAB-MS *m/z*: 468.3836 (Calcd for C₃₁H₄₉O₂N+H: 468.3842); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.35 (1H, d, *J*=5.2 Hz, H-6), 4.73 (1H, ddd, *J*=10.1, 7.9, 5.8 Hz, H-16), 3.51 (1H, m, H-3), 2.46 (1H, d, *J*=10.1 Hz, H-17), 2.43 (2H×2, m, pyrrolidine-2), 2.25 (2H, d, *J*=6.7 Hz, H-26), 1.75 (2H×2, m, pyrrolidine-3), 1.59 (3H, s, 21-CH₃), 1.02 (3H, s, 19-CH₃), 0.93 (3H, d, *J*=6.7 Hz, 27-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 152.0 (C-22), 140.8 (C-5), 121.3 (C-6), 103.3 (C-20), 84.2 (C-16), 71.5 (C-3), 64.2 (C-17), 63.6 (C-26), 55.0 (C-14), 54.4 (pyrrolidine), 50.1 (C-9), 43.2 (C-13), 42.2 (C-4), 39.5 (C-1), 37.2 (C-12), 36.6 (C-10), 34.1 (C-15), 32.8 (C-24), 32.1 (C-7), 32.0 (C-25), 31.6 (C-2), 31.2 (C-8), 23.4 (C-11, pyrrolidine), 21.0 (C-23), 19.4 (C-19), 18.3 (C-27), 13.9 (C-18), 11.6 (C-21).

26-(Piperidin-1-yl)pseudodiosgenin (18) The reaction of **8** (200 mg, 0.38 mmol) with piperidine (130 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The resulting residue was subjected to column chromatography (a gradient of 0–90% acetone in toluene) to give **18** (148 mg; 80% yield) as a powder, mp 170–172 °C. FAB-MS: *m/z* 504 [M+Na]⁺; FAB-MS *m/z*: 482.4003 (Calcd for C₃₂H₅₁O₂N+H: 482.3998); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.34 (1H, d, *J*=5.2 Hz, H-6), 4.72 (1H, ddd, *J*=10.1, 7.6, 5.5 Hz, H-16), 3.51 (1H, m, H-

3), 2.46 (1H, d, *J*=10.1 Hz, H-17), 2.28 (2H×2, m, piperidine-2), 2.11 (1H, dd, *J*=12.2, 6.4 Hz, H-26a), 2.02 (1H, dd, *J*=12.2, 7.9 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.54 (2H×2, m, piperidine-3), 1.41 (2H, m, piperidine-4), 1.02 (3H, s, 19-CH₃), 0.89 (3H, d, *J*=6.4 Hz, 27-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 152.0 (C-22), 140.8 (C-5), 121.2 (C-6), 103.3 (C-20), 84.2 (C-16), 71.5 (C-3), 66.2 (C-26), 64.2 (C-17), 55.0 (C-14, piperidine), 50.0 (C-9), 43.2 (C-13), 42.2 (C-4), 39.5 (C-1), 37.2 (C-12), 36.6 (C-10), 34.1 (C-15), 32.8 (C-24), 32.1 (C-7), 31.5 (C-2), 31.2 (C-8), 29.8 (C-25), 25.9 (piperidine), 24.5 (piperidine), 23.3 (C-11), 21.0 (C-23), 19.4 (C-19), 18.3 (C-27), 13.9 (C-18), 11.6 (C-21).

26-Morpholinopseudodiosgenin (19) The reaction of **8** (200 mg, 0.32 mmol) with morpholine (132 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The resulting residue was purified by recrystallization from acetone to give **19** (100 mg; 54% yield) as a powder, mp 185–187 °C. FAB-MS: *m/z* 484 [M+H]⁺; FAB-MS *m/z*: 484.3787 (Calcd for C₃₁H₄₉O₃N+H: 484.3791); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.34 (1H, d, *J*=5.2 Hz, H-6), 4.72 (1H, ddd, *J*=10.1, 7.9, 5.8 Hz, H-16), 3.67 (2H×2, td, *J*=4.9, 1.5 Hz, morpholine-2), 3.51 (1H, m, H-3), 2.47 (1H, d, *J*=10.1 Hz, H-17), 2.37 (2H×2, m, morpholine-3), 2.15 (1H, dd, *J*=12.2, 6.7 Hz, H-26a), 2.07 (1H, dd, *J*=12.2, 7.6 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.02 (3H, s, 19-CH₃), 0.91 (3H, d, *J*=6.7 Hz, 27-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 151.9 (C-22), 140.9 (C-5), 121.3 (C-6), 103.5 (C-20), 84.2 (C-16), 71.6 (C-3), 67.0 (morpholine), 65.8 (C-26), 64.2 (C-17), 55.0 (C-14), 54.0 (morpholine), 50.1 (C-9), 43.2 (C-13), 42.3 (C-4), 39.5 (C-1), 37.3 (C-12), 36.6 (C-10), 34.1 (C-15), 32.5 (C-24), 32.2 (C-7), 31.6 (C-2), 31.2 (C-8), 29.4 (C-25), 23.2 (C-11), 21.0 (C-23), 19.4 (C-19), 18.1 (C-27), 13.9 (C-18), 11.7 (C-21).

26-(4-Methylpiperazin-1-yl)pseudodiosgenin (20) The reaction of **8** (200 mg, 0.32 mmol) with 1-methylpiperazine (152 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The resulting residue was purified by recrystallization from acetone to give **20** (117 mg; 62% yield) as a powder, mp 207–209 °C. FAB-MS: *m/z* 497 [M+H]⁺; FAB-MS *m/z*: 497.4106 (Calcd for C₃₂H₅₂O₂N₂+H: 497.4107); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.34 (1H, d, *J*=5.2 Hz, H-6), 4.72 (1H, ddd, *J*=10.1, 7.9, 5.8 Hz, H-16), 3.51 (1H, m, H-3), 2.46 (1H, d, *J*=10.1 Hz, H-17), 2.41 (2H×4, m, piperazine), 2.27 (3H, s, N-CH₃), 2.17 (1H, dd, *J*=12.2, 6.4 Hz, H-26a), 2.07 (1H, dd, *J*=12.2, 7.9 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.02 (3H, s, 19-CH₃), 0.89 (3H, d, *J*=6.4 Hz, 27-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 151.9 (C-22), 141.0 (C-5), 121.2 (C-6), 103.5 (C-20), 84.2 (C-16), 71.4 (C-3), 65.3 (C-26), 64.2 (C-17), 55.1 (piperazine), 55.0 (C-14), 53.4 (piperazine), 50.1 (C-9), 46.0 (N-CH₃), 43.2 (C-13), 42.3 (C-4), 39.5 (C-1), 37.3 (C-12), 36.6 (C-10), 34.1 (C-15), 32.6 (C-24), 32.2 (C-7), 31.6 (C-2), 31.2 (C-8), 29.7 (C-25), 23.3 (C-11), 21.0 (C-23), 19.4 (C-19), 18.2 (C-27), 13.9 (C-18), 11.7 (C-21).

26-(1*H*-Imidazol-1-yl)pseudodiosgenin (26) The reaction of **8** (200 mg, 0.38 mmol) with imidazole (104 mg, 1.52 mmol) and K₂CO₃ (525 mg, 3.8 mmol) was performed using standard procedures to obtain a residue. The resulting residue was subjected to column chromatography (a gradient of 0–100% acetone in toluene) to give **26** (59 mg; 33% yield) and **10** (71 mg; 47% yield). Compound **26** was a powder, mp 181–184 °C. FAB-MS: *m/z* 465 [M+H]⁺; FAB-MS *m/z*: 465.3470 (Calcd for C₃₀H₄₄O₂N₂+H: 465.3481); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 7.43 (1H, s, imidazole-2), 7.04 (1H, s, imidazole-4), 6.87 (1H, s, imidazole-5), 5.34 (1H, d, *J*=5.5 Hz, H-6), 4.73 (1H, ddd, *J*=10.1, 8.1, 5.5 Hz, H-16), 3.52 (1H, m, H-3), 3.89 (1H, dd, *J*=13.7, 5.8 Hz, H-26a), 3.67 (1H, dd, *J*=13.7, 7.9 Hz, H-26b), 2.47 (1H, d, *J*=10.1 Hz, H-17), 1.58 (3H, s, 21-CH₃), 1.02 (3H, s, 19-CH₃), 0.87 (3H, d, *J*=6.7 Hz, 27-CH₃), 0.68 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ: 150.8 (C-22), 141.0 (C-5), 137.3 (imidazole), 128.9 (imidazole), 121.0 (C-6), 119.3 (imidazole), 104.2 (C-20), 84.3 (C-16), 71.3 (C-3), 64.0 (C-17), 54.9 (C-14), 53.1 (C-26), 50.0 (C-9), 43.2 (C-13), 42.2 (C-4), 39.4 (C-1), 37.2 (C-12), 36.5 (C-10), 34.3 (C-25), 34.0 (C-15), 32.2 (C-7), 31.5 (C-2), 31.4 (C-24), 31.2 (C-8), 23.0 (C-11), 20.9 (C-23), 19.3 (C-19), 17.2 (C-27), 13.9 (C-18), 11.6 (C-21).

22-((6*R*)-4,5,6,7-Tetrahydro-6-methyl-1*H*-1,2,3-triazepin-4-yl)pseudodiosgenin (30) The reaction of **8** (200 mg, 0.38 mmol) with NaN₃ (99 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The resulting residue was subjected to column chromatography (a gradient of 0–30% ethyl acetate in toluene) to give **30** (50 mg; 43% yield) as a foam. FAB-MS: *m/z* 440 [M+H]⁺; FAB-MS *m/z*: 440.3255 (Calcd for C₂₇H₄₁O₂N₃+H: 440.3277); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ: 5.30 (1H, d, *J*=5.2 Hz, H-6), 4.93 (1H, ddd, *J*=7.6, 6.4, 3.7 Hz, H-16), 4.31 (1H, t, *J*=7.6 Hz, H-23), 3.52 (1H, m, H-3), 3.25 (1H, dd, *J*=11.9, 5.8 Hz, H-26a), 2.09 (1H, dd, *J*=11.9, 7.0 Hz, H-26b), 2.03 (2H, td, *J*=7.6, 1.2 Hz, H-24), 2.00 (1H, d, *J*=6.4 Hz, H-17), 1.79 (1H, m, H-25), 1.54 (3H,

s, 21-CH₃), 1.02 (3H, s, 19-CH₃), 0.96 (3H, d, $J=6.4$ Hz, 27-CH₃), 0.82 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 162.9 (C-22), 140.8 (C-5), 121.2 (C-6), 91.2 (C-23), 84.2 (C-16), 77.6 (C-20), 71.6 (C-3), 66.2 (C-17), 57.4 (C-26), 56.8 (C-14), 49.8 (C-9), 42.2 (C-4), 40.2 (C-13), 39.0 (C-12), 37.2 (C-1), 36.5 (C-10), 34.2 (C-25), 33.1 (C-15), 31.8 (C-7), 31.6 (C-2), 30.9 (C-8), 29.5 (C-24), 21.2 (C-21), 20.3 (C-11), 19.4 (C-19), 17.7 (C-27), 13.2 (C-18).

26-Cyanothiopseudodiosgenone (5) The reaction of **9** (200 mg, 0.38 mmol) with KSCN (148 mg, 1.52 mmol) was performed using standard procedures to obtain a residue. The resulting residue was subjected to preparative HPLC (15% H₂O–acetone) to give **5** (137 mg; 79% yield) as a semisolid. FAB-MS: m/z 476 [M+Na]⁺; FAB-MS m/z : 454.2754 (Calcd for C₂₈H₃₀O₂NS+H: 454.2780); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.72 (1H, s, H-4), 4.75 (1H, ddd, $J=10.1$, 7.9, 5.5 Hz, H-16), 3.03 (1H, dd, $J=12.8$, 5.5 Hz, H-26a), 2.81 (1H, dd, $J=12.8$, 7.6 Hz, H-26b), 2.48 (1H, d, $J=10.1$ Hz, H-17), 1.60 (3H, s, 21-CH₃), 1.20 (3H, s, 19-CH₃), 1.08 (3H, d, $J=6.4$ Hz, 27-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 199.3 (C-3), 170.9 (C-5), 150.8 (C-22), 123.8 (C-4), 112.7 (SCN), 104.2 (C-20), 84.1 (C-16), 64.0 (C-17), 54.1 (C-14), 53.6 (C-9), 43.3 (C-13), 41.2 (C-26), 39.2 (C-12), 38.6 (C-10), 35.6 (C-1), 34.9 (C-8), 33.9 (C-2), 33.2 (C-25), 32.7 (C-6), 32.5 (C-24), 32.2 (C-7 and C-15), 23.0 (C-11), 20.9 (C-23), 18.3 (C-27), 17.3 (C-19), 14.0 (C-18), 11.6 (C-21).

Pseudodiosgenon-25(26)-ene (11) The reaction of **9** (200 mg, 0.38 mmol) with KOCN (62 mg, 0.76 mmol) was performed using standard procedures to obtain a residue. The resulting residue was subjected to preparative HPLC (15% H₂O–acetone) to give **11** (61 mg; 40% yield) as a semisolid. FAB-MS: m/z 417 [M+Na]⁺; FAB-MS m/z : 395.2939 (Calcd for C₂₇H₃₈O₂+H: 395.2951); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.73 (1H, s, H-4), 4.74 (1H, ddd, $J=10.4$, 8.2, 5.8 Hz, H-16), 4.70 (2H, m, H-26), 2.48 (1H, d, $J=10.4$ Hz, H-17), 1.73 (3H, s, 27-CH₃), 1.59 (3H, s, 21-CH₃), 1.20 (3H, s, 19-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 199.3 (C-3), 171.0 (C-5), 151.5 (C-22), 145.2 (C-25), 123.8 (C-4), 110.0 (C-26), 103.5 (C-20), 84.0 (C-16), 64.1 (C-17), 54.1 (C-14), 53.7 (C-9), 43.2 (C-13), 39.3 (C-12), 38.6 (C-10), 35.6 (C-1), 35.1 (C-15), 34.9 (C-8), 33.9 (C-2 and C-24), 32.7 (C-6), 32.2 (C-7), 24.3 (C-11), 22.3 (C-27), 20.9 (C-23), 17.3 (C-19), 14.0 (C-18), 11.5 (C-21).

5 α ,26-Dicyanocolestan-3-one (14) and 5 β ,26-Dicyanocolestan-3-one (15) from **9** The reaction of **9** (200 mg, 0.38 mmol) with NaCN (75 mg, 1.53 mmol) was performed for 12 h as described to obtain a residue. The resulting residue was subjected to preparative HPLC (37.5% H₂O–acetone) to give **14** (31 mg; 18%) and **15** (31 mg; 18%). FAB-MS of **14**: m/z 471 [M+Na]⁺; FAB-MS m/z : 448.3087 (Calcd for C₂₉H₄₀O₂N₂: 448.3090); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 4.75 (1H, ddd, $J=10.4$, 7.9, 5.8 Hz, H-16), 2.54 (1H, d, $J=15.9$ Hz, H-4a), 2.51 (1H, d, $J=10.4$ Hz, H-17), 2.47 (1H, d, $J=15.9$ Hz, H-4b), 2.35 (1H, dd, $J=16.8$, 5.50 Hz, H-26a), 2.24 (1H, dd, $J=16.8$, 7.3 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.15 (3H, s, 19-CH₃), 1.09 (3H, d, $J=6.7$ Hz, 27-CH₃), 0.69 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 206.1 (C-3), 150.9 (C-22), 122.1 (5-CN), 118.8 (26-CN), 104.2 (C-20), 84.2 (C-16), 64.0 (C-17), 53.9 (C-14), 49.3 (C-9), 47.3 (C-5), 47.2 (C-4), 43.5 (C-13), 39.1 (C-12), 37.9 (C-10), 37.1 (C-1), 34.2 (C-6 and C-8), 33.8 (C-2), 33.2 (C-24), 31.4 (C-15), 29.9 (C-25), 28.3 (C-7), 24.3 (C-26), 23.1 (C-11), 21.4 (C-23), 19.3 (C-27), 14.2 (C-18), 12.1 (C-19), 11.6 (C-21). FAB-MS of **15**: m/z 471 [M+Na]⁺; FAB-MS m/z : 448.3085 (Calcd for C₂₉H₄₀O₂N₂: 448.3090); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 4.74 (1H, ddd, $J=10.1$, 7.9, 5.5 Hz, H-16), 2.99 (1H, d, $J=15.9$ Hz, H-4a), 2.50 (1H, d, $J=10.1$ Hz, H-17), 2.38 (1H, d, $J=15.9$ Hz, H-4b), 2.34 (1H, dd, $J=16.8$, 5.2 Hz, H-26a), 2.24 (1H, dd, $J=16.8$, 7.33 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.28 (3H, s, 19-CH₃), 1.08 (3H, d, $J=6.7$ Hz, 27-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 206.8 (C-3), 151.1 (C-22), 122.6 (5-CN), 118.8 (26-CN), 104.0 (C-20), 84.1 (C-16), 64.1 (C-17), 54.4 (C-14), 45.7 (C-5), 44.2 (C-4), 43.3 (C-13), 40.2 (C-9), 39.2 (C-12), 37.2 (C-10), 36.5 (C-1), 33.9 (C-8), 33.8 (C-2), 33.4 (C-6), 33.2 (C-24), 31.1 (C-15), 30.0 (C-25), 26.0 (C-7), 24.4 (C-26), 23.1 (C-11), 21.3 (C-23), 19.6 (C-19), 19.3 (C-27), 14.1 (C-18), 11.6 (C-21).

26-Cyanopseudodiosgenone (13), 5 α ,26-Dicyanocolestan-3-one (14) and 5 β ,26-Dicyanocolestan-3-one (15) from **9** The reaction of **9** (200 mg, 0.38 mmol) with NaCN (75 mg, 1.53 mmol) was performed for 2.5 h to obtain a residue. The resulting residue was subjected to preparative HPLC (15% H₂O–acetone) to give **13** (54 mg; 33% yield) and a mixture. The mixture was subjected to preparative HPLC (DOCOSIL-B, 37.5% H₂O–acetone) to give **14** (20 mg, 12%) and **15** (30 mg, 17%). FAB-MS of **13**: m/z 444 [M+Na]⁺; FAB-MS m/z : 422.3048 (Calcd for C₂₈H₃₀O₂N+H: 422.3059); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.73 (1H, s, H-4), 4.74 (1H, ddd, $J=10.1$, 7.9, 5.5 Hz, H-16), 2.48 (1H, d, $J=10.1$ Hz, H-17), 2.35 (1H, dd, $J=16.8$, 5.2 Hz, H-26a), 2.24 (1H, dd, $J=16.8$, 7.3 Hz, H-26b),

1.59 (3H, s, 21-CH₃), 1.20 (3H, s, 19-CH₃), 1.09 (3H, d, $J=6.7$ Hz, 27-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 199.5 (C-3), 171.0 (C-5), 150.8 (C-22), 123.9 (C-4), 118.7 (CN), 104.2 (C-20), 84.2 (C-16), 64.0 (C-17), 54.1 (C-14), 53.7 (C-9), 43.3 (C-13), 39.3 (C-12), 38.6 (C-10), 35.7 (C-1), 35.0 (C-8), 33.9 (C-2), 33.1 (C-24), 32.7 (C-6 and C-15), 32.2 (C-7), 29.9 (C-25), 24.3 (C-26), 23.1 (C-11), 20.9 (C-23), 19.2 (C-27), 17.4 (C-19), 14.0 (C-18), 11.6 (C-21).

26-Diethylaminopseudodiosgenone (21) The reaction of **9** (200 mg, 0.38 mmol) with diethyl amine (112 mg, 1.53 mmol) was performed as described to obtain a residue. The resulting residue was purified by column chromatography (a gradient of 0–90% acetone in toluene) to give **21** (115 mg; 64% yield) as a semisolid. FAB-MS: m/z 490 [M+Na]⁺; FAB-MS m/z : 468.3858 (Calcd for C₃₁H₄₉O₂N+H: 468.3842); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.73 (1H, s, H-4), 4.72 (1H, ddd, $J=10.1$, 7.6, 5.5 Hz, H-16), 2.48 (4H, CH₂×2, q , $J=7.3$ Hz, NCH₂CH₃), 2.47 (1H, d, $J=10.1$ Hz, H-17), 2.24 (1H, dd, $J=12.5$, 5.8 Hz, H-26a), 2.11 (1H, dd, $J=12.5$, 7.6 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.20 (3H, s, 19-CH₃), 0.99 (6H, CH₃×2, t , $J=7.0$ Hz, NCH₂CH₃), 0.90 (3H, d, $J=6.4$ Hz, 27-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 199.2 (C-3), 170.9 (C-5), 152.1 (C-22), 123.7 (C-4), 103.0 (C-20), 83.9 (C-16), 64.0 (C-17), 60.0 (C-26), 54.1 (C-14), 53.6 (C-9), 47.3 (NCH₂CH₃), 43.2 (C-13), 39.2 (C-12), 38.5 (C-10), 35.6 (C-1), 34.9 (C-8), 33.9 (C-2), 33.8 (C-24), 32.7 (C-6), 32.3 (C-7), 31.0 (C-25), 23.3 (C-11), 20.8 (C-23), 18.2 (C-27), 17.3 (C-19), 13.9 (C-18), 11.5 (C-21 and NCH₂CH₃).

26-(Pyrrolidin-1-yl)pseudodiosgenone (22) The reaction of **9** (200 mg, 0.38 mmol) with pyrrolidine (108 mg, 1.52 mmol) was performed as described to obtain a residue. The resulting residue was purified by column chromatography (a gradient of 0–90% acetone in toluene) to give **22** (48 mg; 27% yield) as a semisolid. FAB-MS: m/z 466 [M+H]⁺; FAB-MS m/z : 466.3680 (Calcd for C₃₁H₄₇O₂N+H: 466.3686); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.73 (1H, s, H-4), 4.37 (1H, ddd, $J=10.1$, 7.9, 5.8 Hz, H-16), 2.47 (1H, d, $J=10.1$ Hz, H-17), 2.40 (2H×2, m, pyrrolidine-2), 2.28 (2H, d, $J=7.3$ Hz, H-26), 1.77 (2H×2, m, pyrrolidine-3), 1.59 (3H, s, 21-CH₃), 1.20 (3H, s, 19-CH₃), 0.95 (3H, d, $J=6.7$ Hz, 27-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 199.3 (C-3), 171.0 (C-5), 152.2 (C-22), 123.8 (C-4), 103.2 (C-20), 84.0 (C-16), 64.1 (C-17), 63.6 (C-26), 54.8 (pyrrolidine), 54.1 (C-14), 53.7 (C-9), 43.3 (C-13), 39.3 (C-12), 38.6 (C-10), 35.7 (C-1), 35.0 (C-8), 33.9 (C-2 and C-15), 32.8 (C-24), 32.7 (C-6), 32.2 (C-7), 32.0 (C-25), 23.4 (C-11, pyrrolidine), 20.9 (C-23), 18.3 (C-27), 17.3 (C-19), 14.0 (C-18), 11.6 (C-21).

26-Piperidinopseudodiosgenone (23) The reaction of **9** (200 mg, 0.38 mmol) with piperidine (130 mg, 1.52 mmol) was performed as described to obtain a residue. The resulting residue was purified by column chromatography (a gradient of 0–90% acetone in toluene) to give **23** (146 mg; 79% yield) as a semisolid. FAB-MS: m/z 502 [M+Na]⁺; FAB-MS m/z : 480.3838 (Calcd for C₃₂H₄₉O₂N+H: 480.3842); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.72 (1H, s, H-4), 4.72 (1H, ddd, $J=10.1$, 7.9, 5.8 Hz, H-16), 2.47 (1H, d, $J=10.1$ Hz, H-17), 2.29 (2H×2, m, piperidine-2), 2.11 (1H, dd, $J=11.9$, 6.4 Hz, H-26a), 2.02 (1H, dd, $J=11.9$, 8.2 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.55 (2H×2, m, piperidine-3), 1.41 (2H, m, piperidine-4), 1.20 (3H, s, 19-CH₃), 0.89 (3H, d, $J=6.7$ Hz, 27-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 199.3 (C-3), 171.0 (C-5), 152.3 (C-22), 123.9 (C-4), 103.2 (C-20), 84.0 (C-16), 66.3 (C-26), 64.1 (C-17), 55.0 (piperidine), 54.2 (C-14), 53.7 (C-9), 43.3 (C-13), 39.3 (C-12), 38.6 (C-10), 35.7 (C-1), 35.0 (C-8), 34.0 (C-15), 33.9 (C-2), 32.8 (C-6 and C-24), 32.3 (C-7), 29.9 (C-25), 26.0 (piperidine), 24.6 (piperidine), 23.3 (C-11), 21.0 (C-23), 18.4 (C-27), 17.4 (C-19), 14.0 (C-18), 11.6 (C-21).

26-Morpholinopseudodiosgenone (24) The reaction of **9** (200 mg, 0.38 mmol) with morpholine (132 mg, 1.52 mmol) was performed as described to obtain a residue. The resulting residue was purified by column chromatography (a gradient of 0–50% acetone in toluene) to give **24** (135 mg; 73% yield) as a semisolid. FAB-MS: m/z 482 [M+Na]⁺; FAB-MS m/z : 482.3618 (Calcd for C₃₁H₄₇O₃N+H: 482.3635); ¹H-NMR (CDCl₃) (only assignable signals were listed) δ : 5.73 (1H, s, H-4), 4.72 (1H, ddd, $J=10.1$, 7.9, 5.8 Hz, H-16), 3.69 (2H×2, t , $J=4.0$ Hz, morpholine-2), 2.83 (2H×2, t , $J=4.0$ Hz, morpholine-3), 2.47 (1H, d, $J=10.1$ Hz, H-17), 2.17 (1H, dd, $J=12.2$, 4.9 Hz, H-26a), 2.07 (1H, dd, $J=12.2$, 7.6 Hz, H-26b), 1.59 (3H, s, 21-CH₃), 1.20 (3H, s, 19-CH₃), 0.91 (3H, d, $J=6.4$ Hz, 27-CH₃), 0.72 (3H, s, 18-CH₃); ¹³C-NMR (CDCl₃) δ : 199.2 (C-3), 170.8 (C-5), 152.0 (C-22), 123.7 (C-4), 103.2 (C-20), 83.9 (C-16), 66.9 (morpholine), 65.7 (C-26), 64.0 (C-17), 54.1 (C-14), 53.9 (morpholine), 53.6 (C-9), 43.2 (C-13), 39.2 (C-12), 38.5 (C-10), 35.6 (C-1), 34.9 (C-8), 33.8 (C-2 and C-15), 32.6 (C-6 and C-24), 32.2 (C-7), 29.3 (C-25), 23.3 (C-11), 20.8 (C-23), 18.0 (C-27), 17.3 (C-19), 13.9 (C-18), 11.5 (C-21).

26-(4-Methylpiperazin-1-yl)pseudodiosgenone (25) The reaction of **9** (200 mg, 0.38 mmol) with 1-methylpiperazine (152 mg, 1.52 mmol) was performed as described to obtain a residue. The resulting residue was purified by column chromatography (a gradient of 0–100% acetone in toluene) to give **25** (115 mg; 61% yield) as a semisolid. FAB-MS: m/z 495 $[M+Na]^+$; FAB-MS m/z : 495.3942 (Calcd for $C_{32}H_{50}O_2N_2+H$: 495.3951); 1H -NMR ($CDCl_3$) (only assignable signals were listed) δ : 5.73 (1H, s, H-4), 4.72 (1H, ddd, $J=10.4, 7.9, 5.8$ Hz, H-16), 2.47 (1H, d, $J=10.4$ Hz, H-17), 2.42 (2H \times 4, m, piperazine), 2.30 (3H, s, NCH_3), 2.17 (1H, dd, $J=12.2, 5.5$ Hz, H-26a), 2.08 (1H, dd, $J=12.2, 7.9$ Hz, H-26b), 1.59 (3H, s, 21- CH_3), 1.20 (3H, s, 19- CH_3), 0.90 (3H, d, $J=6.4$ Hz, 27- CH_3), 0.72 (3H, s, 18- CH_3); ^{13}C -NMR ($CDCl_3$) δ : 199.3 (C-3), 170.9 (C-5), 152.0 (C-22), 123.7 (C-4), 103.2 (C-20), 83.9 (C-16), 65.2 (C-26), 64.0 (C-17), 55.0 (piperazine), 54.1 (C-14), 53.6 (C-9), 53.2 (piperazine), 45.9 (NCH_3), 43.2 (C-13), 39.2 (C-12), 38.5 (C-10), 35.6 (C-1), 34.9 (C-8), 33.9 (C-15), 33.8 (C-2), 32.7 (C-24), 32.5 (C-6), 32.2 (C-7), 29.6 (C-25), 23.2 (C-11), 20.8 (C-23), 18.1 (C-27), 17.2 (C-19), 13.9 (C-18), 11.5 (C-21).

26-(1H-Imidazol-1-yl)pseudodiosgenone (27) The reaction of **9** (200 mg, 0.38 mmol) with imidazole (104 mg, 1.52 mmol) and K_2CO_3 (525 mg, 3.8 mmol) was performed as described to obtain a residue. The resulting residue was purified by column chromatography (a gradient of 0–100% acetone in toluene) to give **27** (32 mg; 18% yield) as a semisolid and **11** (41 mg; 27% yield). FAB-MS of **27**: 463 $[M+H]^+$; FAB-MS m/z : 463.3322 (Calcd for $C_{30}H_{42}O_2N_2+H$: 463.3325); 1H -NMR ($CDCl_3$) (only assignable signals were listed) δ : 7.44 (1H, s, imidazole-2), 7.05 (1H, s, imidazole-4), 6.88 (1H, s, imidazole-5), 5.73 (1H, s, H-4), 4.73 (1H, ddd, $J=10.1, 7.9, 5.8$ Hz, H-16), 3.89 (1H, dd, $J=13.7, 5.8$ Hz, H-26a), 3.68 (1H, dd, $J=13.7, 7.9$ Hz, H-26b), 2.48 (1H, d, $J=10.1$ Hz, H-17), 1.58 (3H, s, 21- CH_3), 1.20 (3H, s, 19- CH_3), 0.88 (3H, d, $J=6.4$ Hz, 27- CH_3), 0.71 (3H, s, 18- CH_3); ^{13}C -NMR ($CDCl_3$) δ : 199.4 (C-3), 170.9 (C-5), 151.0 (C-22), 137.4 (imidazole), 129.1 (imidazole), 123.8 (C-4), 119.3 (imidazole), 104.0 (C-20), 84.1 (C-16), 64.0 (C-17), 54.1 (C-14), 53.7 (C-9), 53.1 (C-26), 43.3 (C-13), 39.2 (C-12), 38.6 (C-10), 35.7 (C-1), 35.0 (C-8), 34.3 (C-25), 33.9 (C-2 and C-15), 32.7 (C-6), 32.2 (C-7), 31.4 (C-24), 23.0 (C-11), 20.9 (C-23), 17.3 (C-27), 17.2 (C-19), 14.0 (C-18), 11.6 (C-21).

26-Azidopseudodiosgenone (31) The reaction of **9** (200 mg, 0.38 mmol) with NaN_3 (99 mg, 1.52 mmol) was performed as described to obtain a residue. The resulting residue was subjected to preparative HPLC (15% H_2O -acetone) to give **31** (103 mg; 61% yield) as a semisolid. FAB-MS: m/z 438 $[M+H]^+$; FAB-MS m/z : 438.3109 (Calcd for $C_{27}H_{39}O_2N_3+H$: 438.3121); 1H -NMR ($CDCl_3$) (only assignable signals were listed) δ : 5.72 (1H, s, H-4), 4.74 (1H, ddd, $J=10.1, 7.9, 5.8$ Hz, H-16), 3.23 (1H, dd, $J=11.9, 5.5$ Hz, H-26a), 3.11 (1H, dd, $J=11.9, 7.0$ Hz, H-26b), 2.49 (1H, d, $J=10.1$ Hz, H-17), 1.59 (3H, s, 21- CH_3), 1.20 (3H, s, 19- CH_3), 0.97 (3H, d, $J=6.7$ Hz, 27- CH_3), 0.72 (3H, s, 18- CH_3); ^{13}C -NMR ($CDCl_3$) δ : 199.2 (C-3), 170.8 (C-5), 151.3 (C-22), 123.7 (C-4), 103.7 (C-20), 84.0 (C-16), 63.9 (C-17), 57.5 (C-26), 54.0 (C-14), 53.6 (C-9), 43.2 (C-13), 39.2 (C-12), 38.5 (C-10), 35.6 (C-1), 34.9 (C-8), 33.8 (C-2 and C-15), 32.9 (C-25), 32.6 (C-6), 32.2 (C-7), 23.0 (C-11), 20.8 (C-23), 17.4 (C-27), 17.3 (C-19), 13.9 (C-18), 11.5 (C-21).

2-((6R)-4,5,6,7-Tetrahydro-6-methyl-1H-1,2,3-triazepin-4-yl)pseudodiosgenone (32) from 31 A solution of **31** (200 mg, 0.46 mmol) in DMF (20 ml) was heated at 120 °C for 20 h, and the mixture was evaporated. The resulting residue was subjected to preparative HPLC (30% H_2O -acetone) to give **32** (62 mg; 31% yield) as a semisolid. FAB-MS: m/z 438 $[M+H]^+$; FAB-MS m/z : 438.3117 (Calcd for $C_{27}H_{39}O_2N_3+H$: 438.3121); 1H -NMR ($CDCl_3$) (only assignable signals were listed) δ : 5.73 (1H, s, H-4), 4.92 (1H, ddd, $J=10.1, 6.1, 3.7$ Hz, H-16), 4.30 (1H, t, $J=7.6$ Hz, H-23), 3.23 (1H, dd, $J=11.9, 5.8$ Hz, H-26a), 3.08 (1H, dd, $J=11.9, 7.0$ Hz, H-26b), 2.03 (2H, t, $J=7.6$ Hz, H-24), 1.99 (1H, d, $J=6.4$ Hz, H-17), 1.79 (1H, m, H-25), 1.52 (3H, s, 21- CH_3), 1.18 (3H, s, 19- CH_3), 0.95 (3H, d, $J=6.7$ Hz, 27- CH_3), 0.84 (3H, s, 18- CH_3); ^{13}C -NMR ($CDCl_3$) δ : 199.4 (C-3), 170.8 (C-5), 162.7 (C-22), 123.8 (C-4), 91.1 (C-23), 83.8 (C-16), 77.3 (C-20), 66.1 (C-17), 57.3 (C-26), 55.9 (C-14), 53.3 (C-9), 40.1 (C-13), 38.6 (C-12), 38.4 (C-10), 35.5 (C-1), 34.5 (C-8), 34.1 (C-25), 33.8 (C-2), 32.8 (C-15), 32.6 (C-6), 31.8 (C-7), 29.4 (C-24), 21.1 (C-21), 20.1 (C-11), 17.6 (C-27), 17.2 (C-19), 13.1 (C-18).

Cytotoxic Activities, Cell Lines and Culture The human colorectal carcinoma cell line (HCT 116, ATCC No. CCL-247) and human hepatoma cell line (Hep G2, No. RCB0459) were purchased from Dainippon Pharmaceutical Co. LTD (Osaka, Japan) and RIKEN Cell Bank (Tsukuba, Japan), respectively. Dulbecco's Modified Eagle's medium (DMEM), McCoy's 5A medium, fetal bovine serum (FBS) and the penicillin–streptomycin mixture were purchased from Wako Pure Chemical Industries Ltd. (Osaka, Japan),

Sigma (MO, U.S.A.), Biosource International (CA, U.S.A) and Bio Whittaker (ND, U.S.A), respectively. The HCT 116 cells and Hep G2 cells were maintained in McCoy's 5A medium and in DMEM, respectively. Each medium was supplemented with 10% FBS and the penicillin–streptomycin mixture (100 U/ml penicillin and 100 mg/ml streptomycin) at 37 °C in a humidified atmosphere containing 5% CO_2 .

Assay Procedure The ability of the drug to inhibit cellular growth was determined using the MTT assay.²²⁾ The cytotoxicity of test drugs was assessed as previously described.²⁸⁾ Each experiment was performed in duplicate wells, and all experiments involved a control (DMSO only). The drug treatments were performed separately 3 times. Data represent mean \pm S.D. values.

Antibacterial and Antifungal Activities Test Organisms: *Bacillus subtilis* IFO 3134, *Staphylococcus aureus* IFO 12732, *Escherichia coli* IFO 3972, *Pseudomonas aeruginosa* IFO 13275, *Candida albicans* IFO 1594 and *Aspergillus niger* IFO 9455 were purchased from the Institute for Fermentation (Osaka, Japan).

Assay for Antibacterial Activities: Assays for activity against gram-positive (*B. subtilis*, *S. aureus*) and gram-negative (*E. coli*, *P. aeruginosa*) bacteria were carried out by the micro dilution method according to the NCCLS documents M7-A5.²⁵⁾ The bacterial strains were maintained on trypticase soy agar at 27 °C. The test compounds in DMSO were diluted with Mueller Hinton II broth (cation adjusted) to give serial 2-fold dilutions in the range 500–3.9 μ g/ml. The presence of DMSO in the assay medium (maximum concentration 5%) did not affect the growth of the bacteria. The inoculum was standardized by adjusting to 1×10^6 CFU/ml for each bacterium. The minimal inhibitory concentration (MIC) was estimated after incubation at 37 °C for 24 h. Each experiment was performed in duplicate well and drug treatments were performed separately 3 times.

Assay for Antifungal Activities: Antifungal assays were carried out by the micro dilution method according to the NCCLS documents M27-A for *Candida* and H38-P for *Aspergillus*.^{26,27)} Strains of *A. niger* and *C. albicans* were maintained on potato dextrose agar and Sabouraud dextrose agar, respectively, at 27 °C. Test compounds in DMSO were diluted with RPMI 1640 medium to give serial 2-fold dilutions in the range 500–3.9 μ g/ml. The growth of tested fungus was not affected by the presence of DMSO (maximum concentration 5%). The inoculum was standardized by adjusting to 8×10^4 CFU/ml for *A. niger* and to 2.5×10^3 CFU/ml for *C. albicans*. MIC was estimated after incubation at 27 °C for 48 h. Each experiment was performed in duplicate wells and drug treatments were performed separately 3 times.

References

- Hayashi K., Wada K., Mitsunashi H., Bando H., Takase M., Terada S., Koide Y., Aiba T., Narita T., Mizuno D., *Chem. Pharm. Bull.*, **28**, 1954–1958 (1980).
- Mita A., Shida R., Kasai N., Shoji J., *Biomedicine*, **31**, 223–227 (1979).
- Cuellar M. J., Giner R. M., Recio M. C., Just M. J., Manez S., Cerds M., Hostettmann K., Rios J.-L., *Nat. J. Prod.*, **60**, 1158–1160 (1997).
- Ito M., Nakashima H., Baba M., Shigeta S., Yamamoto N., *Igaku no Ayumi*, **141**, 427–428 (1987).
- Ito M., Nakashima H., Baba M., Shigeta S., Yamamoto N., *Chem. Abstr.*, **107**, 89374y (1987).
- Saito S., Kuroda K., Hayashi Y., Nagamura Y., Nishida K., Ishiguro I., *Chem. Pharm. Bull.*, **39**, 2333–2339 (1991).
- Saito S., Sumita S., Kanda Y., Sasaki Y., *Tetrahedron Lett.*, **33**, 7381–7384 (1992).
- Saito S., Sasaki Y., Kuroda K., Hayashi Y., Sumita S., Nagamura Y., Nishida K., Ishiguro I., *Chem. Pharm. Bull.*, **41**, 539–543 (1993).
- Saito S., Ebashi J., Sumita S., Furumoto T., Nagamura Y., Nishida K., Ishiguro I., *Chem. Pharm. Bull.*, **41**, 1395–1401 (1993).
- Saito S., Sumita S., Kanda Y., Sasaki Y., *Chem. Pharm. Bull.*, **42**, 1016–1027 (1994).
- Saito S., Sumita S., Furumoto T., Ebashi J., Nagamura Y., Ishiguro I., *Eur. J. Med. Chem.*, **29**, 455–470 (1994).
- Saito S., Nagase S., Kawase M., Nagamura Y., *Eur. J. Med. Chem.*, **31**, 557–574 (1996).
- Saito S., Furumoto T., Ochiai M., Hosono A., Hoshino H., Haraguchi U., Ikeda R., Shimada N., *Eur. J. Med. Chem.*, **31**, 365–381 (1996).
- Beaton J. M., Spring F. S., *J. Chem. Soc.*, **1955**, 3126–3129.
- Quan H.-J., Koyanagi J., Ohmori K., Uesato S., Tsuchido T., Saito S., *Eur. J. Med. Chem.*, **37**, 659–669 (2002).
- Uhle F. C., *J. Org. Chem.*, **27**, 2797–2799 (1962).

- 17) Uhle F. C., *J. Org. Chem.*, **32**, 792—797 (1967).
- 18) Uhle F. C., *J. Org. Chem.*, **32**, 1596—1601 (1967).
- 19) Shionoya M., Jimbo T., Kitagawa M., Soga T., Tohgo A., *Cancer Sci.*, **94**, 459—466 (2003).
- 20) Lee G., Piqette-Miller M., *Can. J. Physiol. Pharmacol.*, **79**, 876—884 (2001).
- 21) Takara K., Sakaeda T., Tanigawara Y., Nishiguchi K., Ohmoto N., Horinouchi M., Komada F., Ohnishi N., Yokoyama T., Okumura K., *Eur. J. Pharm. Sci.*, **16**, 159—165 (2002).
- 22) Mosmann T., *J. Immunol. Methods*, **65**, 55—63 (1983).
- 23) Tsuchido T., Takeuchi H., Kawahara H., Obata H., *J. Ferment. Bioeng.*, **78**, 183—187 (1994).
- 24) Tsuchido T., Yasunaga K., Matsumura Y., Oku K., *Biocontrol Sci.*, **1**, 61—63 (1996).
- 25) National committee for clinical laboratory standards. 2000, M7-A5.
- 26) National committee for clinical laboratory standards. 1997, M27-A.
- 27) National committee for clinical laboratory standards. 1998, M38-P.
- 28) Jin G.-Z., Quan H.-J., Koyanagi J., Takeuchi K., Miura Y., Komada F., Saito S., *Cancer Lett.*, **218**, 15—20 (2005).