

Discrimination of Sayamakaori and Yabukita Which Are Original Plant Source of Japanese Green Tea and Identification of Specific Compounds for the Former by Nuclear Magnetic Resonance Metabolomics Techniques Combined with Isolation by Chromatography Methods

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Abstract

Camellia sinensis leaves are one of the most economically important crops, and various tea cultivars are available in Japan. In the present study, we compared the constituents of *Camellia sinensis* var. *sinensis* cv. Yabukita (“Yabukita”), a standard tea cultivar, and those of *C. sinensis* var. *sinensis* cv. Sayamakaori (“Sayamakaori”), grown in Saitama Prefecture, using proton nuclear magnetic resonance (¹H-NMR) metabolomics technology. Principal component analysis and hierarchical cluster analysis of ¹H-NMR spectroscopic data of the MeOH extracts of *C. sinensis* leaves revealed differences in the metabolic profiles of each tea cultivar. Chromatographic separation based on ¹H-NMR spectral data showed that kaempferol 3-O-β-D-glucopyranosyl-(1 → 3)-α-L-rhamnopyranosyl-(1 → 6)-β-D-glucopyranoside was a distinct component of “Sayamakaori”. This compound can be used as marker for the identification of “Sayamakaori” and the maintenance of its quality.

Keywords

tea cultivars (*Camellia sinensis*), Sayamakaori, ¹H-NMR metabolomics, flavonoids, kaempferol 3-O-β-D-glucopyranosyl-(1 → 3)-α-L-rhamnopyranosyl-(1 → 6)-β-D-glucopyranoside

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Introduction

The tea plant *Camellia sinensis* is one of the most economically important crops in Japan and other countries. The three main types of tea can be classified according to fermentation (fermentation here means an oxidation process) as follows: green (unfermented), oolong (semi-fermented), and black (fermented). Flavan-3-ols, phenolic acids, purine alkaloids, condensed tannins, hydrolyzable tannins, saponins, flavonols, and their glycosides are secondary metabolites in fresh tea leaves. Particularly, flavan-3-ol is a basic skeleton of (–)-epicatechin (EC), (–)-epicatechin gallate (ECG), (–)-epigallocatechin (EGC), and (–)-epigallocatechin gallate (EGCG), which are well known green tea ingredients.^{1,2} These components have anticancer, anti-cardiovascular, neuroprotective, antioxidant, hepatoprotective, anti-obesity, antidiabetic, antibacterial, and antiviral effects.^{3,4} Studies on volatile components have also been conducted, and more than 600 volatile components have been identified to date.⁵ Although various tea cultivars are cultivated in Japan, almost all of these have been bred from *C. sinensis* var. *sinensis* cv. Yabukita (“Yabukita”). “Yabukita” accounts for over 70% of the cultivation area in Japan. Some compositional studies regarding the differences between cultivars have been conducted, and it has been concluded that the differences were due to

differences in the concentrations of common components.^{6–8} *C. sinensis* var. *sinensis* cv. Sayamakaori (“Sayamakaori”), originating from Saitama Prefecture, Japan, is said to have a strong young shoot aroma that differs from that of “Yabukita” and has a bitter taste and high yield. However, the components that characterize “Sayamakaori” have not yet been clarified. Characteristic compounds can be used to distinguish between species and contribute to quality control. Furthermore, these marker compounds may be useful for counterfeit detection as protecting crop brands such as tea is important.

Metabolite fingerprinting approaches aided by nuclear magnetic resonance (NMR) spectroscopy provide valuable metabolite signatures for complex plant extracts. The analysis per

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sample is fast, no derivatization is required, and specific chemical entities can be identified using this method.^{9–11} Some studies have used NMR-based metabolomic analysis to characterize plant cultivars and genera and identify the characteristic components of these cultivars. In addition, NMR metabolomics is known to be suitable for identifying the differentiable compounds of some cultivars.^{12–14} In this study, we determined the characteristic components of the tea cultivar “Sayamakaori” using NMR metabolome analysis and multivariate statistical analyses, including principal component (PCA) and hierarchical cluster analyses (HCA).

Results and Discussion

A total of 23 samples were used in this study (Table 1). The samples were macerated with methanol to obtain the corresponding extracts. After thoroughly drying in a vacuum, the extracts were dissolved in CD₃OD for ¹H-NMR analysis. NMR spectra were analyzed using PCA and HCA. PCA was depicted as a score plot and consisted of two composite variables: principal component PC1 (with the largest variance of the data) and PC3 (with the third-largest variance of the data). The samples were divided into two groups based on PCA score plots (Figure 1). These two groups were related to

Table 1. Tea Samples Used in this Experiment.

Sample no.	Tea cultivars	Year the tree was planted
1	Sayamakaori	1982
2		1993
3		2010
4		2014
5		2015
6		2015
7		2017
8		2018
9		2019
10	Yabukita	2012, Deep pruning 2019
11		1971
12		1971
13		1980
14		1992
15		1993
16		2010
17		2014
18		2015
19		2015
20		2017
21		2018
22		2019
23	2012, Deep pruning 2019	

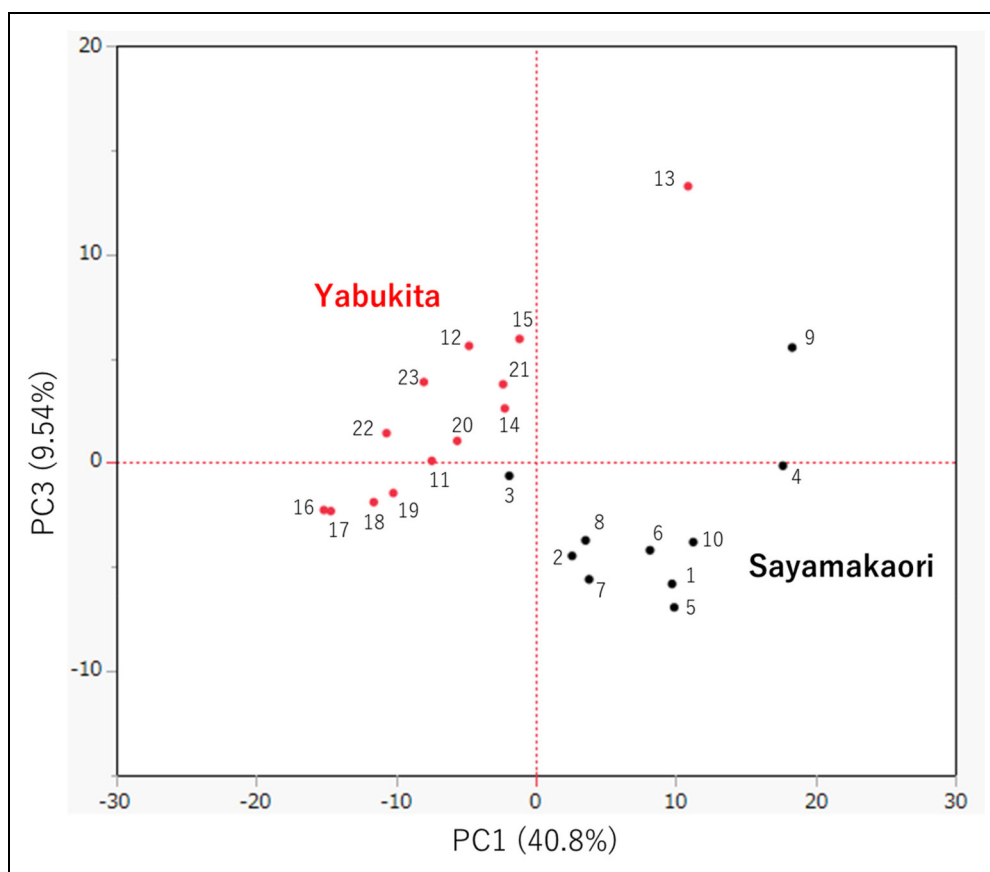


Figure 1. PCA score plot based on ¹H-NMR spectra of 23 tea samples. Numerals in the score plot represent samples in Table 1.

the variety and were distinguished into the “Sayamakaori” and “Yabukita” groups. In addition, the HCA constellation dendrogram of *C. sinensis* leaves was divided into two groups (Figure 2). These findings indicate that the components of the tea cultivars differ from each other. Next, we identified an $^1\text{H-NMR}$ signal specific to the leaves of “Sayamakaori.” As shown in Figure 3, these signals were selected as specific signals for “Sayamakaori” in the loading plot of the PCA. Among these signals, δ 8.02 was focused on because it appeared independently without overlapping with other signals in the $^1\text{H-NMR}$ spectrum. The signal was statistically specific for “Sayamakaori” ($P < 0.0001$), as shown in Figure 4. Furthermore, a signal was observed at δ 8.02 for “Sayamakaori” by direct comparison of $^1\text{H-NMR}$ spectra (Figure 5). Therefore, we attempted to isolate the compound whose signal appeared at δ 8.02 using chromatographic methods. The “Sayamakaori” methanol extract was fractionated based on the specific $^1\text{H-NMR}$ signals using a Diaion HP-20 polymeric adsorbent (Mitsubishi Chemical Industries Ltd, Chiyoda, Tokyo, Japan) and eluted with water and 30%, 50%, and 70% aqueous MeOH to obtain the corresponding fractions. Thereafter, polymerized tannins were removed from the 70% methanolic elute by passing it through an open silica gel

column. The elute was concentrated under reduced pressure, and the residue was purified by medium-pressure liquid chromatography with octadecyl silica gel to obtain Fr.1 to Fr.240. The specific signals for “Sayamakaori” were observed in Fr. 92-95, using $^1\text{H-NMR}$ spectroscopy. Furthermore, $^1\text{H-NMR}$ spectra indicated that these fractions (Fr. 92-95) contained only one pure component. Therefore, these fractions were combined and designated as compound **1**. In addition, using detailed 2D-NMR analysis and comparison of the $^1\text{H-}$ and $^{13}\text{C-NMR}$ and MS data, compound **1** was identified as kaempferol 3-*O*- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside by comparing NMR and MS data with literature ($^{13}\text{C-NMR}$ spectral data was shown in Supplemental Table 1).¹⁵ The NMR proton signals specific for “Sayamakaori” observed at δ 8.02 (d, $J = 8.96$ Hz) were assigned to H-2' (H-6') of compound **1** (Figure 6). In a previous study, Monobe et al measured the contents of flavonol glycosides in 37 green tea samples using LC-MS analysis.¹⁶ According to their results, the content of compound **1** in “Sayamakaori” was higher than that in “Yabukita” (Saymakaori:92.8 $\mu\text{g/mL}$, Yabukita:20.8 $\mu\text{g/mL}$, each in dried leaf). However, the authors did not even touch this point other than showing the results in the paper. This was not possible

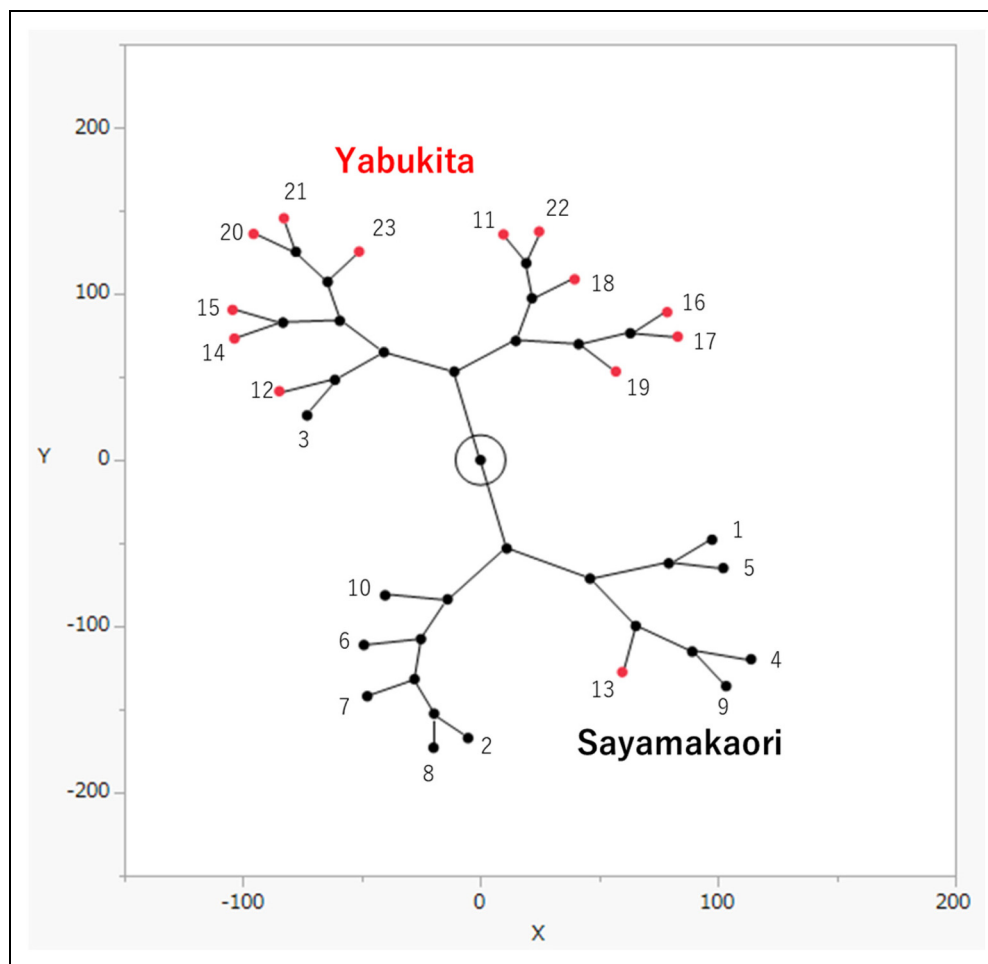


Figure 2. HCA constellation dendrogram derived from $^1\text{H-NMR}$ spectral data. Numerals in the score plot represent samples in Table 1.

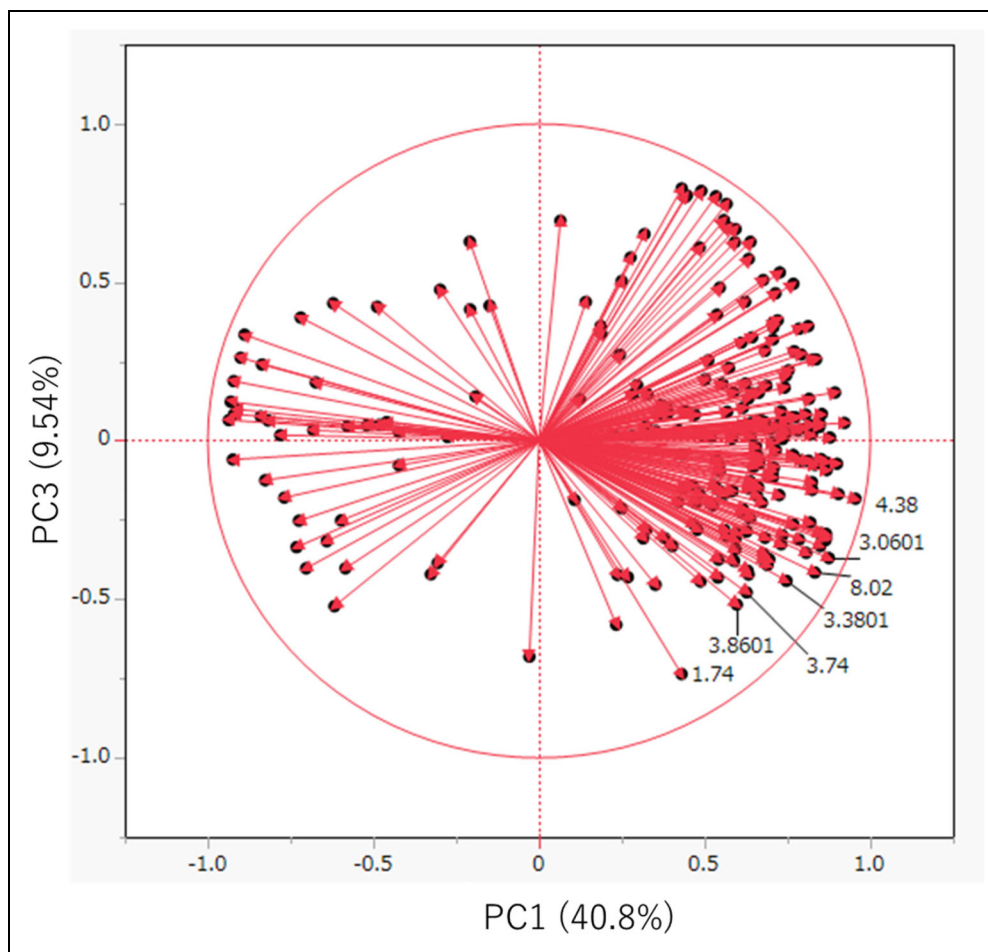


Figure 3. Loading plot of PCA. Numerals in the loading plot represent ^1H -NMR chemical shifts.

without NMR metabolomic analysis. These results indicated that compound **1** could be an important metabolite for discrimination between the “Sayamakaori” and “Yabukita”. This compound has been reported to exert a protective effect against D-galactosamine-induced liver injury. It has also been shown to exhibit lipid accumulation inhibitory activity in HepG2 cells.¹⁷ By the way, this experiment results indicated that samples of “Sayamakaori” were categorized as two groups on the PCA score plot. Therefore, we intend to clarify why “Sayamakaori” is divided into two groups on the PCA score plot in future research.

Material and Methods

The Reagents

Methanol- d_4 (CD_3OD) was purchased from Kanto Chemical Co. (Chuo-ku, Tokyo, Japan).

Tea Samples

Fresh green tea (*C. sinensis* var. *sinensis* cv. Sayamakaori and *C. sinensis* var. *sinensis* cv. Yabukita) leaves (one bud and two

leaves) were obtained from the The Green Tea Laboratory of Saitama Prefectural Agriculture and Forestry Research Center, Iruma, Saitama, on May 7, 2021. All tea leaves were dried at 23 °C for 2 weeks. The specimens were maintained at the Laboratory of Natural Products and Phytochemistry, Department of Pharmaceutical Sciences, Faculty of Pharmacy and Pharmaceutical Sciences, Josai University, Japan.

Instrumentation

^1H - and ^{13}C -NMR spectra were recorded at 400 MHz on an AVANCE NEO 400 (Bruker Corporation, Billerica, MA, USA) operating at 400 MHz for ^1H and 100 MHz for ^{13}C . MS was performed on a JMS-700 double-focusing magnetic sector mass spectrometer (JEOL, Tachikawa, Tokyo, Japan).

Extraction

The tea samples were extracted with MeOH for 24 h at 23 °C, after which the extract was evaporated to obtain the corresponding residue. The MeOH extracts were dissolved in CD_3OD at a concentration of 10 mg/mL for ^1H -NMR spectral measurements.

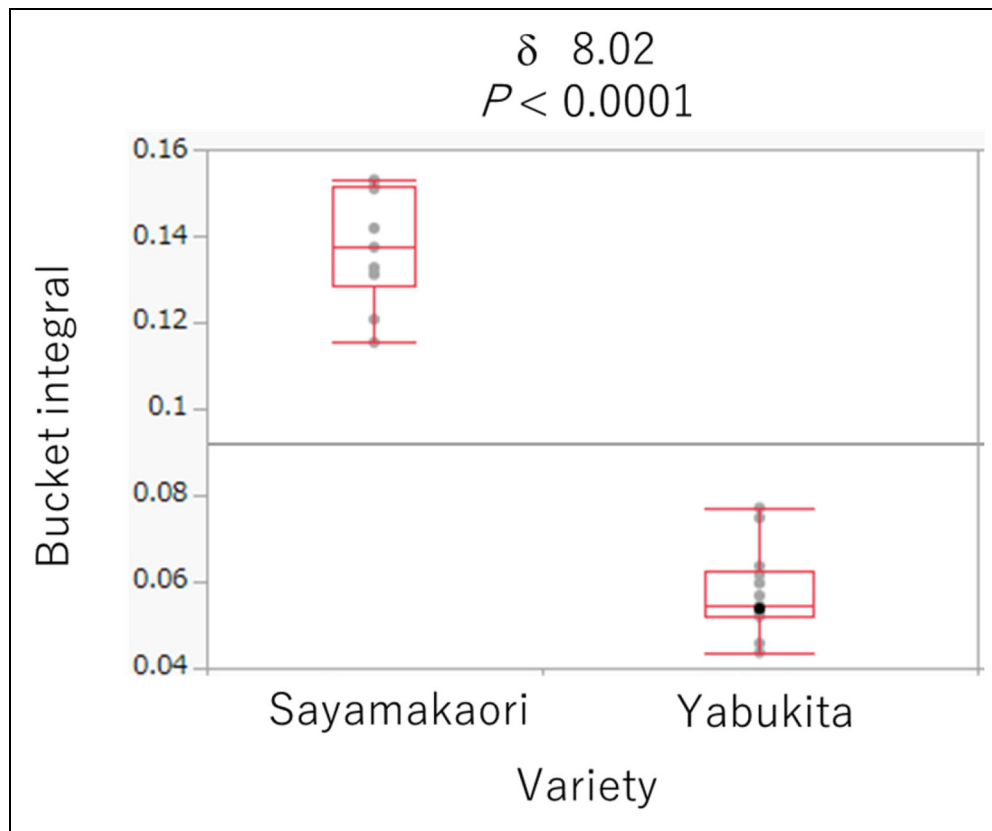


Figure 4. Box plots of the bucket integral at δ 8.02 of “Sayamakaori” and “Yabukita.”

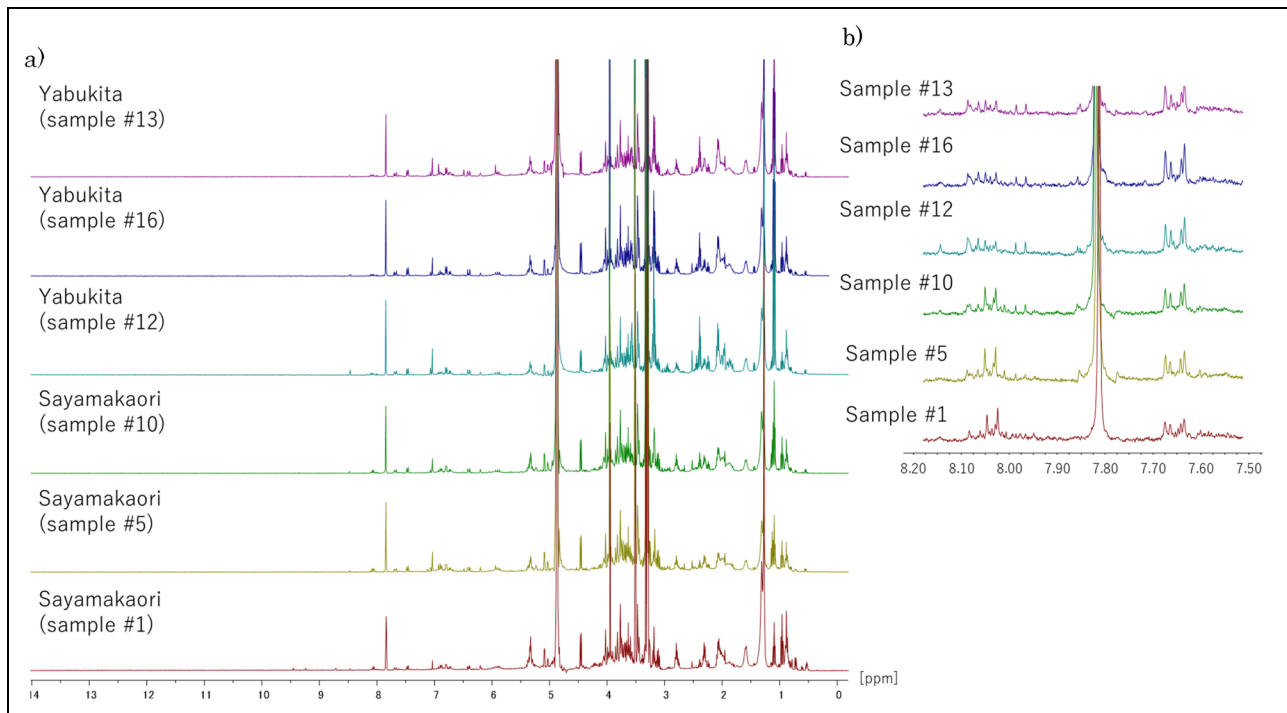


Figure 5. Comparison of ^1H -NMR spectra of representative tea cultivars. (a) Overview, (b) Enlarged view.

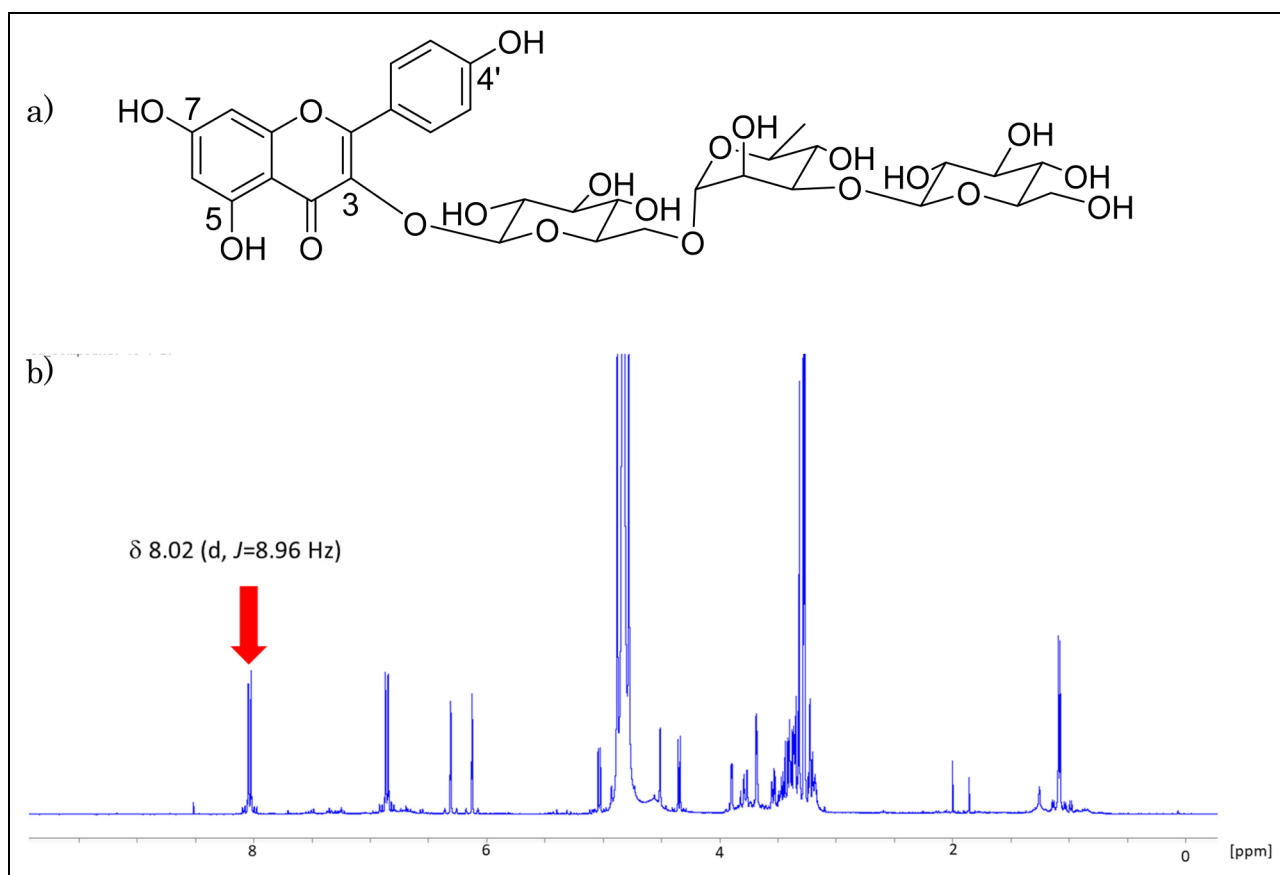


Figure 6. Chemical structure of compound **1** (a) and the ¹H-NMR spectrum of compound **1** (b).

NMR Spectra Measurements of Green Tea Extracts

One-dimensional NMR spectra were recorded at 20 °C with solvent signals as internal references. Each spectrum consisted of 65 536 complex data points with a spectral width of 8196.722 Hz. The spectra were obtained over 16 scans with an acquisition time of 4.0 s and a relaxation delay of 1.00 s per scan. The flip angle of the detection pulse was set to 30°.

NMR Data Reduction Procedures and Pattern Recognition Analysis

Each NMR spectrum was divided into 250 regions between 0.00 and 10.00 ppm (0.04 ppm width). Segments of each spectral region were integrated; the integration regions from 1.12 to 1.24 ppm, 3.20 to 3.36 ppm, and 4.76 to 4.92 ppm containing solvent and water signals were removed from the data leaving 243 regions for all data. The remaining integrals for each spectrum were normalized by summing up 100 integrals to correct for concentration differences among tea extracts. Spectral processing was performed using ALICE2 for Metabolome (version 5.0; JEOL). Multivariate analyses, such as PCA and HCA, were conducted using JMP Pro 16 (SAS Institute Inc., Cary, NC, USA). The variables were standardized with a mean of 0 and a standard

deviation of 1. For HCA, Euclidean distances and Ward's method were used to establish the dendrograms. A Mann-Whitney U test was used to compare the differences in the integral value at δ 8.02 between "Sayamakaori" and "Yabukita," and a *P*-value < 0.0001 was considered statistically significant.

Isolation and Identification of Kaempferol-3-O-β-D-glucopyranosyl-(1 → 3)-α- L-Rhamnopyranosyl-(1 → 6)-β-D-Glucopyranoside

A large amount of "Sayamakaori" was extracted again to isolate a specific compound for this variate. Methanol extracts prepared by extracting a large amount of "Sayamakaori" were fractionated guided by ¹H-NMR signals identified as specific for "Sayamakaori." The leaves of "Sayamakaori" (400 g) were extracted with MeOH for 24 h at room temperature, and the same procedure was repeated twice for 24 h. The MeOH extracts were combined and concentrated to obtain 108 g of residue. Since a specific signal on δ 8.07 was observed in this residue by ¹H-NMR measurement, 100 g of the residue was subjected to a Diaion HP-20 polymeric adsorbent (Mitsubishi Chemical Industries Ltd) and eluted with water and 30%, 50%, and 70% aqueous MeOH to obtain the corresponding fractions. (Water fraction, 26.7 g; 30% aqueous MeOH fraction,

31.7 g; 50% aqueous MeOH fraction, 21.0 g; 70% aqueous MeOH fraction, 3.7 g; MeOH fraction, 10.7 g). The 70% aqueous MeOH fraction (3.4 g) was loaded onto an open silica gel column (ϕ 50 × 420 mm) and separated by elution with CHCl₃: H₂O = 5:1. The eluate (200 mg) was purified by preparative MPLC using an ULTRA PACK ODS-SM-50C size C (ϕ 37 × 300 mm; Yamazen Corp., Osaka, Japan). Preparative conditions were eluted in an isocratic system (MeOH: H₂O = 1:1, 6 mL/min) to obtain Fr.1–Fr.240. Each fraction was collected at 6 mL/tube. Fr. 92–95 contained a specific component of “Sayamakaori.” Each fraction was combined with Compound 1 (4.71 mg), which was identified as kaempferol 3-O- β -D-glucopyranosyl-(1 → 3)- α -L-rhamnopyranosyl-(1 → 6)- β -D-glucopyranoside. This structure was confirmed by comparing the NMR and MS data with those in the literature.

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.


Ethical Approval

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Statement of Human and Animal Rights

This article does not include contain any studies on human or animal subjects.

Informed Consent

There are no human subjects in this article and informed consent is not applicable.

Supplemental Material

Supplemental material for this article is available online.

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