

難代謝性フッ素化脂肪酸の生体作用の多様性 と作用点における分子識別機構の解明

(研究課題番号 09672247)

平成9年度～平成11年度科学研究費補助金
(基盤研究 C(2))

研究成果報告書

平成12年 3 月

研究代表者 川嶋 洋一

(城西大学薬学部 教授)

難代謝性フッ素化脂肪酸の生体作用の多様性 と作用点における分子識別機構の解明

(研究課題番号 09672247)

平成9年度～平成11年度科学研究費補助金
(基盤研究 C(2))

研究成果報告書

平成12年 3 月

研究代表者 川嶋 洋一

(城西大学薬学部 教授)

目 次

はしがき	1
研究組織・研究経費	2
研究発表	2
研究成果	
ペルフルオロデカン酸によるラット肝オレイン酸生成促進作用	5
生体内ペルフルオロカルボン酸のガスクロマトグラフィーに よる定量	11
高速液体クロマトグラフィーによるペルフルオロ脂肪酸の定量	19
ペルフルオロオクタン酸によるラット肝グリセロ脂質代謝の 変動	26
炭素鎖長の異なる種々のフッ素化脂肪酸によるラット肝ペル オキシソーム β 酸化酵素誘導能の比較	41
フッ素化脂肪酸によるラット肝脂肪蓄積作用	55

は し が き

ペルフルオロ脂肪酸 (PFCA) は脂肪酸の炭素結合水素がすべてフッ素に置換した化合物である。PFCAは通常の脂肪酸にはない特殊な性格をもっている。すなわち、表面張力が極めて弱いため水ばかりでなく、ほとんどの有機溶媒中でも界面活性作用を示す。また、C-F結合はC-H結合に比べて結合エネルギーが大きい化学的にもきわめて安定である。このような性質を利用して撥水剤、撥油剤、消火剤などに広く利用されている。この化合物の生体毒性は非常に低いと考えられている。しかし、化学工場労働者の血液からPFCAの一種であるペルフルオロオクタン酸が検出されるという報告も出されている。生体に蓄積した微量の化学物質が生体に影響をあたえるという環境ホルモンの例は、目立った毒性がないからといって、蓄積性の高い化学物質を使用することの危険性を示している。

本研究代表者らはこれまでにペルフルオロ脂肪酸の生理作用について研究を重ね、これらの化合物がげっ歯類肝臓の脂質代謝や薬物代謝に広く影響を与えることを明らかにしてきた。その過程で、炭素鎖長の異なるPFCAはときとして生体への作用が異なることが明らかになってきた。このことは炭素1つの違いを生体は何らかの形で認識していることを示している。本研究ではPFCAの炭素鎖長と生体作用の関係を系統的に調べ、生体がPFCAのどのような構造を認識するのかについて知見を得ることを目的とした。さらに、生体作用の差がどのような機構で生ずるのか、すなわち、生体のPFCA分子識別の機構を解明する手がかりを得たいと考えた。本研究によって、PFCAの生体作用として脂質代謝酵素系への影響が新たに明らかになった。さらに、PFCAの生体作用の違いを統一的に理解する上で重要な知見が得られたので、これらを報告させて頂く。

本研究に対する文部省からの化学研究補助金の交付に対して深く感謝する次第である。

研究組織

研究代表者： 川嶋 洋一 (城西大学薬学部・教授)

研究分担者： 工藤 なをみ (城西大学薬学部・助手)
大谷 武司 (城西大学薬学部・講師)
三浦 裕晃 (マルホ株式会社
研究開発本部中央研究所・主任研究員)

研究経費

平成 9 年度	2, 2 0 0 千円
平成 1 0 年度	5 0 0 千円
平成 1 1 年度	6 0 0 千円
計	3, 3 0 0 千円

研究発表

(1) 学会誌等

1. Yamamoto, A. and Kawashima, Y., Perfluorodecanoic acid enhances the formation of oleic acid in rat liver. Biochem. J., 325, 429-434 (1997)
2. Kudo, N., Bandai, N. and Kawashima, Y., Quantitation of perfluorocarboxylic acids by gas-liquid chromatography in rat tissues. Toxicol. Lett., 99, 183-190 (1998)
3. Ohya, T., Kudo, N., Suzuki, E. and Kawashima, Y., Determination of perfluorinated carboxylic acids by high-performance chromatography in biological samples., J. Chromatogr. B, 720, 1-7 (1998)
4. Kudo, N., Mizuguchi, H. and Kawashima, Y., Alterations by perfluorooctanoic acid of glycerolipid metabolism in rat liver., Chemico-Biol. Interact., 118, 69-83 (1999)
5. Kudo, N., Bandai, N., Suzuki, E., Katakura, M. and Kawashima, Y., Induction by perfluorinated fatty acids with different carbon chain length of peroxisomal β -oxidation in the liver of rats., Chemico-Biol. Interact., 124, 119-132 (2000)

(2) 口頭発表

1. 工藤なをみ、坂大直樹、川嶋洋一
ペルフルオロカルボン酸によるラットの脂肪肝の誘導
第23回環境トキシコロジーシンポジウム（東京）平成9年10月
2. 工藤なをみ、坂大直樹、川嶋洋一
ペルフルオロカルボン酸によるラットの脂肪肝誘発機構
日本薬学会第118年会（京都）平成10年3月
3. 大谷武司、工藤なをみ、川嶋洋一
蛍光検出-HPLCによるペルフルオロカルボン酸の定量
日本薬学会第118年会（京都）平成10年3月
4. 鈴木恵理春、工藤なをみ、川嶋洋一
ペルフルオロカルボン酸のマウス肝ペルオキシソーム増殖作用
日本薬学会第118年会（京都）平成10年3月
5. 工藤なをみ、鈴木恵理春、川嶋洋一
炭素鎖長の異なるペルフルオロカルボン酸の生体残留性の違い
第24回環境トキシコロジーシンポジウム（京都）平成10年10月
6. 鈴木恵理春、工藤なをみ、川嶋洋一
ラットにおける炭素鎖長の異なるペルフルオロ脂肪酸の排泄の違い
第42回日本薬学会関東支部会平成10年10月
7. 中川和也、工藤なをみ、川嶋洋一
四塩化炭素およびペルフルオロデカン酸による肝脂肪蓄積と魚油摂取の効果
日本薬学会第119年会（徳島）平成11年3月
8. 大森耕太郎、工藤なをみ、川嶋洋一
ペルフルオロオクタン酸のラットにおける生体作用の性差
日本薬学会第119年会（徳島）平成11年3月

9. 工藤 なをみ、鈴木 恵理春、川嶋洋一
ラットにおける炭素鎖長の異なるペルフルオロ脂肪酸の排泄の違い
日本薬学会第119年会（徳島） 平成11年3月
10. 大森耕太郎、工藤なをみ、川嶋洋一
種々のフッ素化脂肪酸の排泄経路の比較検討
第43回日本薬学会関東支部会 平成11年10月
11. 中川和也、工藤なをみ、川嶋洋一
ペルフルオロデカン酸による肝脂肪蓄積機構の検討
第43回日本薬学会関東支部会 平成11年10月
12. 工藤なをみ、大森耕太郎、野城理絵、川嶋洋一
フッ素化脂肪酸の体内残留性の比較検討
第25回環境トキシコロジーシンポジウム（名古屋） 平成11年10月
13. 片倉賢紀、工藤なをみ、川嶋洋一
性ホルモンによるペルフルオロオクタン酸の尿中排泄調節機構
日本薬学会第120年会（岐阜）発表予定 平成12年3月
14. 内藤佳奈、藤兼裕子、工藤なをみ、川嶋洋一
ペルフルオロカルボン酸によるstearyl-CoA不飽和化酵素誘導とリン脂質
アシル基組成への影響
日本薬学会第120年会（岐阜）発表予定 平成12年3月
15. 野城理絵、工藤なをみ、川嶋洋一
長鎖フッ素化脂肪酸の排泄促進の検討
日本薬学会第120年会（岐阜）発表予定 平成12年3月
16. 工藤なをみ、中川和也、川嶋洋一
ペルフルオロデカン酸による肝脂肪蓄積機構の解析
日本薬学会第120年会（岐阜）発表予定 平成12年3月

Induction of hepatic triglyceride accumulation by perfluorinated fatty acids in rat liver

Naomi Kudo, Hiroaki Miura*, Kohtaro, Ohmori, Naoki Bandai and Yoichi Kawashima#

Faculty of Pharmaceutical Sciences, Josai University and *Maruho Research and Laboratories

#, To whom correspondence should be addressed:

Dr. Yoichi Kawashima

Faculty of Pharmaceutical Sciences, Josai University, Keyaki-dai, Sakado, Saitama 350-0295, Japan

Phone: +81 (492) 71-7676

Fax: +81 (492) 71-7984

E-mail address: ykawash@josai.ac.jp

Introduction

Perfluorinated fatty acids (PFCAs), in which all hydrogens are substituted by fluorine, are commercially used as lubricants, wetting agents, corrosion inhibitors, and fire extinguishers. The effects of PFCAs on biological systems have been studied by several investigators. When rats were treated with perfluorooctanoic acid (PFOA, a PFCA with 8 carbon atoms), hepatomegaly and peroxisomal proliferation were observed. The induction of various enzymes involved in lipid metabolism and drug metabolism were also reported in the livers of rats. In addition of the above effects, perfluorodecanoic acid (PFDA), a PFCA having 10 carbon atoms, caused a significant lipid accumulation in rat liver. This raises the question that the induction of hepatic lipid accumulation is a specific effect of PFDA among various PFCAs. There is a sex-related difference of the induction of peroxisomal β -oxidation by PFOA, whereas the effect of PFDA was equally observed in male and female rats. It is not known whether sex-related difference is observed in effect of PFDA on the levels of hepatic lipids. In this context, we compared the effects on hepatic level of triglyceride (TG) between PFCAs having 7-10 carbon atoms in both male and female rats.

Materials and Methods

Materials

Perfluoroheptanoic acid (PFHA), perfluorooctanoic acid (PFOA) and perfluorodecanoic acid (PFDA) were purchased from Aldrich Japan (Tokyo, Japan). Perfluorononanoic acid (PFNA) was obtained from Lancaster Synthesis (Lancashire, UK). Methylheptadecanoate and triheptadecanoin was purchased from Nu-Chek-Prep Inc. (MN, USA). Testosterone propionate was purchased from Wako pure Chemicals Inc. (Osaka, Japan). All other chemicals used were of analytical grade.

Animals

Male and female Wistar rats aged 5 weeks old were purchased from SLC Inc. (Hamamatsu, Japan). After acclimatization for 1 week, rats were received intraperitoneal injections with PFHA, PFOA, PFNA or PFDA once a day for 5 days. PFCAs were dissolved into propyleneglycol:water (1:1, v/v) after neutralization with equimolar NaOH. All rats were received a dosing volume of 1 mL/ kg body weight.

Some of male rats (24-26 day-old) were castrated. Four days after castration, rats were administered with testosterone propionate dissolving into corn oil (10 mg/ kg body weight), or vehicle alone once every two days until being killed. These rats were administered with PFNA or PFDA at a dose of 20 mg / kg body weight as described above.

Twentyfour hours after final injection with PFCAs, rats were killed by decapitation under light ether anesthesia. Livers were quickly excised, perfused with

ice-cold 0.9% NaCl, and then homogenized with 9 volumes of 0.25 M sucrose/ 1 mM EDTA / 10 mM Tris-HCl (pH 7.4).

Histopathological examination

Livers were excised under light ether anesthesia from the rats which have been received intraperitoneal injections of PFDA (20 mg/ kg body weight) or vehicle alone once a day for 5 days. Liver was fixed in neutral-buffered 10% formalin. Fixed specimens were embedded in paraffin, and their sections were stained with Hematoxylin-Eosin for light microscopy. For discrimination of lipid droplets by Oil Red O staining, frozen sections were made with fixed specimens using cryostat (CM-3000, Leica, Germany)

Lipid analysis

Total lipid was extracted from liver homogenates after the addition of triheptadecanoin as an internal standard, according to the method of Bligh and Dyer. Triglyceride was separated by TLC on Silica gel G plates (Merck, Darmstadt, Germany), which were developed with hexane: diethyl ether: acetic acid (80:30:1, v/v/v). The spots corresponding to triglyceride were visualized by spraying 0.005% primuline in 80% acetone (w/v), scraped off from the plates and then transferred to glass tubes. TG was extracted from the silica with 5 mL of chloroform/ methanol/ 0.1 N HCl (4:4:1, v/v/v) twice. Added 3 mL of 0.1 N HCl to the combined extract and then lower layer was transferred to a new tube. Solvent was dried down under

nitrogen stream, 0.5 M sodium methoxide was added to the tubes to prepare fatty acid methyl esters. GLC (Simadzu GC 14A) was employed for the analysis of fatty acid with SpelcowaxTM 10 (0.53 mm ID x 30 m).

Determination of perfluorinated fatty acids

After adding an internal standard, PFCAs were extracted from liver homogenates as an ion pair with tetrabutylammonium, derivitized with 3-bromoacetyl-7-methoxycoumarin (BrAMC) and quantified by HPLC with a fluorescent detection according to the method of Ohya, et al. (). PFHA was used as an internal standard for the determination of PFNA and PFDA, and perfluorohexanoic acid was used for the determination of PFHA and PFOA, respectively.

Statistical analysis

Analysis of variance was used to test the significance of differences between various doses of a PFCA, between different PFCAs at a same dose, and between male and female, castrated male and testosterone –treated male rats which had been castrated. Where Differences were significant, the statistical significance between any two means was determined using Sheffe's multiple range test. Statistical significance between male and female rats was analyzed by Student's t-test or Welch's F-test for two means.

Results and Discussion

Effects of PFCAs on the level of hepatic triglyceride

The level of hepatic TG was compared between the rats being treated with vehicle, PFHA, PFOA, PFNA or PFDA (Figure 1). Treatment with PFDA and PFNA increased hepatic level of TG, whereas PFHA and PFOA did not alter it in male rats. The level of hepatic TG was increased 6-fold by PFDA treatment in male rats (20 mg/ kg) on the basis of g liver. In female rats, only PFDA caused a significant increase in hepatic level of TG. When rats were administered with PFNA or PFDA at doses over 10 mg / kg body weight, weight loss and less appetite were significantly observed. Hepatic TG level of the rats which were restricted in food intake to those consumed by the rats received PFDA (20 mg/ kg) was not significantly different from vehicle treated rats. Therefore, an increase in the level of hepatic TG is not due to restricted food intake in PFCA-treated rats. The accumulation of hepatic TG by PFDA and PFNA (in male rats) was observed in a dose-dependent manner (Figure 2).

Pathological findings

Light microscopy revealed that fine lipid droplets accumulated into hepatocytes, which were found diffusely entire lobule in PFDA-treated male rats (20 mg/ kg B.W., Figure 3). Occasionally, necrotic lesions were recognized focally in lobules by the staining with Hematoxylin and Eosin.

Alterations of fatty acid composition in hepatic lipids

Treatment with PFDA increased all fatty acids except for 20:5(n-3) on the basis of gram liver (Table 1). An increase in 18:1 was greater than those in other fatty acids, therefore, percentage of 18:1 was significantly increased upon PFDA treatment. In phospholipid fraction, composition of oleic acid was also increased. The specific increase in oleic acid is thought to be due to an increase in the production of oleic acid which is catalyzed by stearoyl-coA desaturase.

Effects of hormonal status on hepatic TG accumulation by PFNA

Whether sex-related difference of PFNA-induced TG accumulation in rat liver correlate with hormonal status was determined. Castration significantly reduced hepatic TG level in PFNA-treated male rats, which was close to that of female rats those have been treated with PFNA at the same condition as male. By contrast, treatment of castrated rats with testosterone restored the level of hepatic TG (Figure 4). By contrast to PFNA, there was no significant difference of hepatic TG levels between male and female rats when treated with PFDA. Castration and testosterone treatment did not affect hepatic TG level in PFDA-treated rats. We and other investigators have reported sex-related difference in the induction of peroxisomal β -oxidation by PFOA in rat liver. Such difference was not observed in the case of PFNA. Therefore, this is the first report that describes sex-related difference in the effects of PFNA in rats.

Correlation between the levels of PFCA and TG in rat liver

To explore the reason why sex-related difference was observed in hepatic TG level but not in the activity of peroxisome β -oxidation, correlation with hepatic concentrations of PFCA was examined. We have previously described that a PFCA with longer carbon chain length more accumulates in rat liver. In this context, it is possible that higher accumulation of PFCA cause higher TG accumulation in rat liver. Figure 3 demonstrates positive correlation between the levels of TG and PFCA in the liver regardless of their carbon chain length. It is noteworthy that correlation coefficient between hepatic PFCA concentration and hepatic TG level was significantly different from that between hepatic PFCA concentration and the activity of peroxisomal β -oxidation. Hepatic PFCA concentration required to cause a significant TG accumulation is thought to be 250 $\mu\text{g/g}$ liver. This concentration is higher than those required to induce peroxisomal β -oxidation. This suggests that PFCA caused the induction of peroxisomal β -oxidation and the accumulation of hepatic TG by independent mechanisms.

In summary, we reported that PFCA caused an increase in hepatic TG level, which was dependent on the concentration of hepatic PFCA regardless of their carbon chain length. Sex-related difference of the effect was observed in rats. PFCA seems to cause hepatic TG accumulation by the mechanisms independent on the induction of peroxisomal β -oxidation.

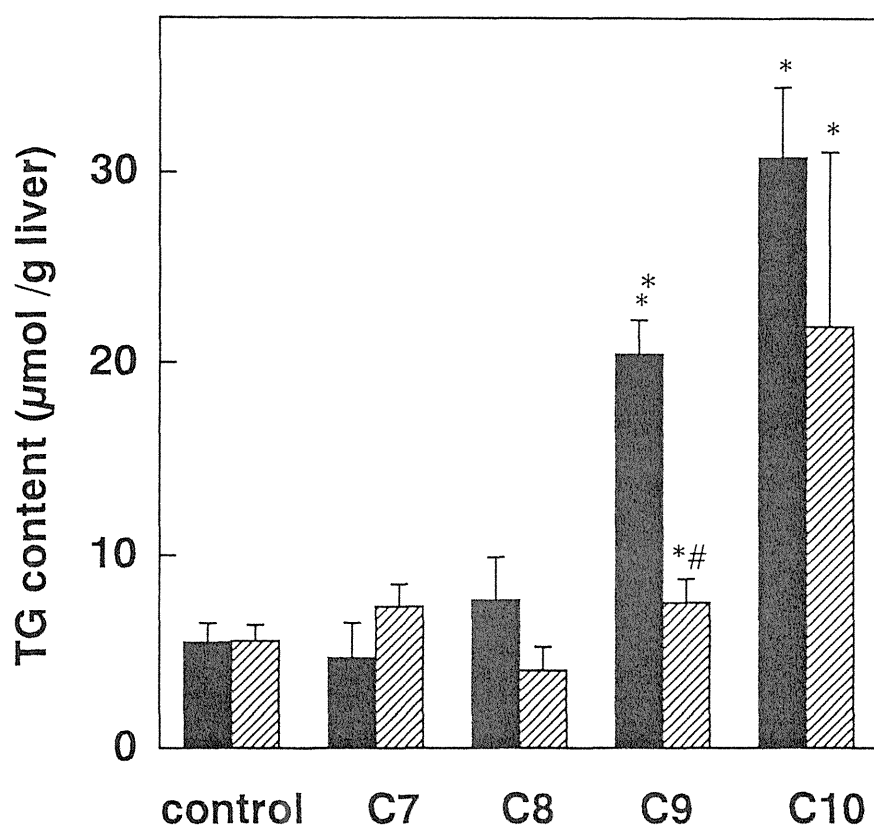


Figure 1 Effects of various PFCAs on the level of hepatic triglyceride in male and female rats

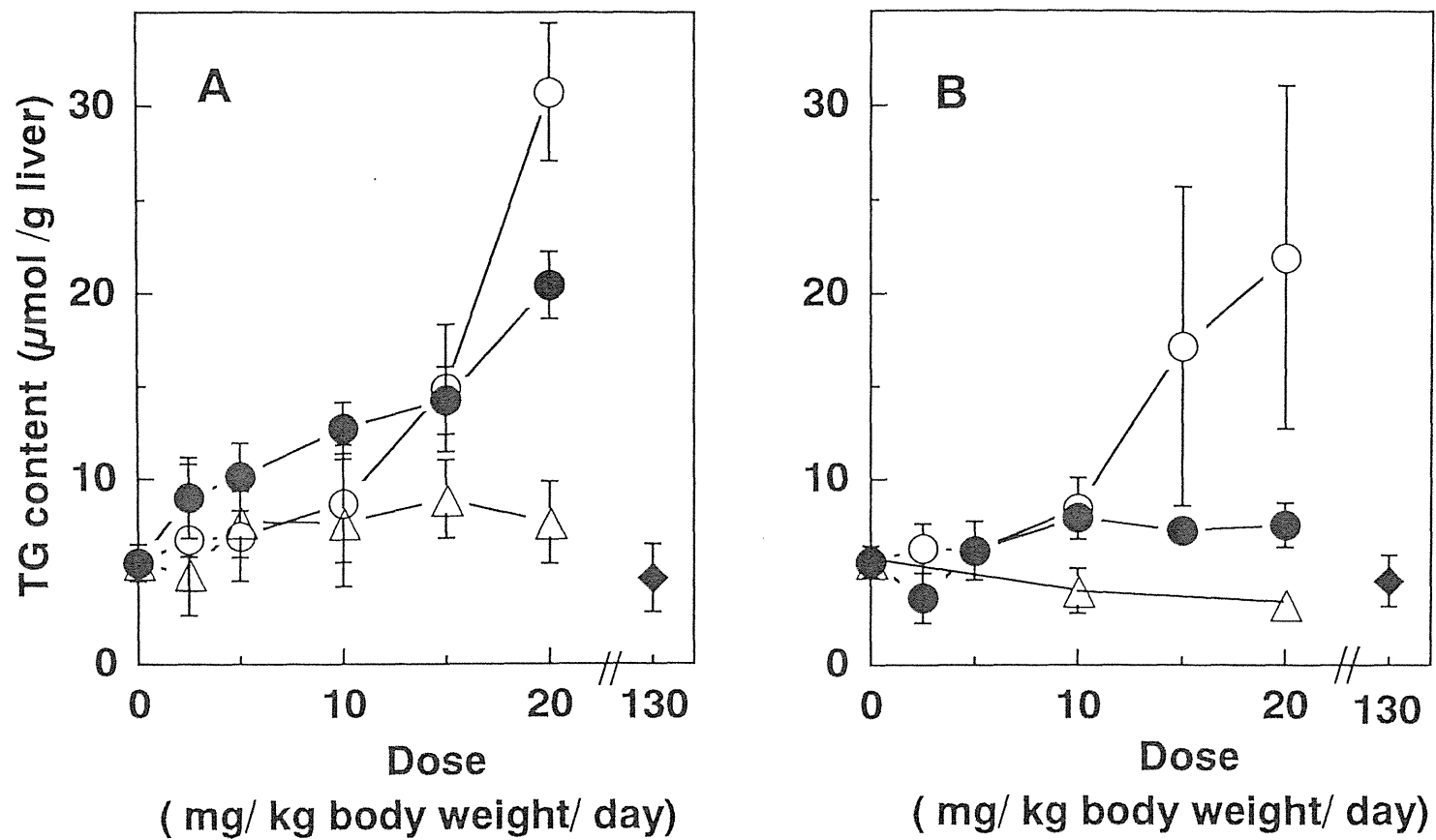
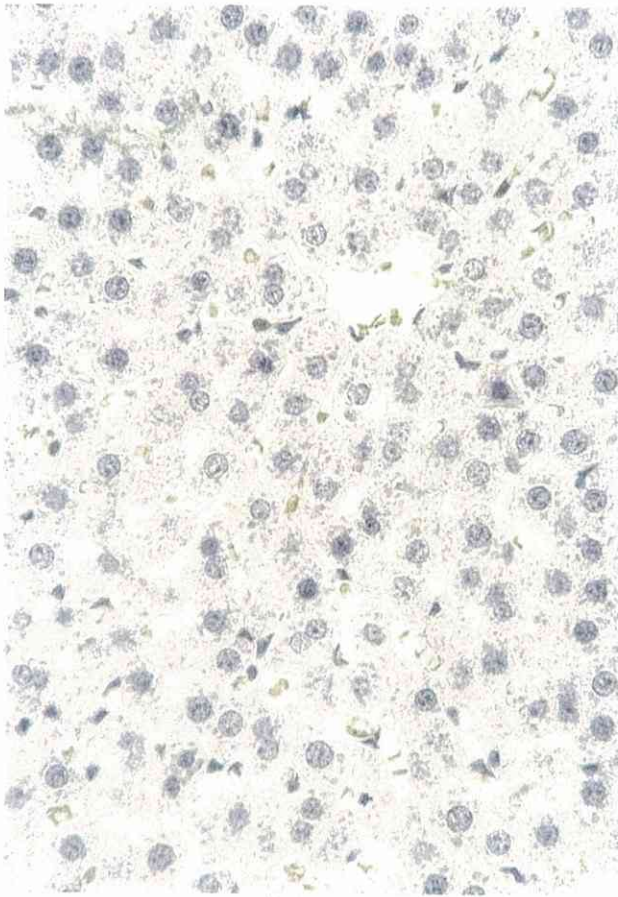
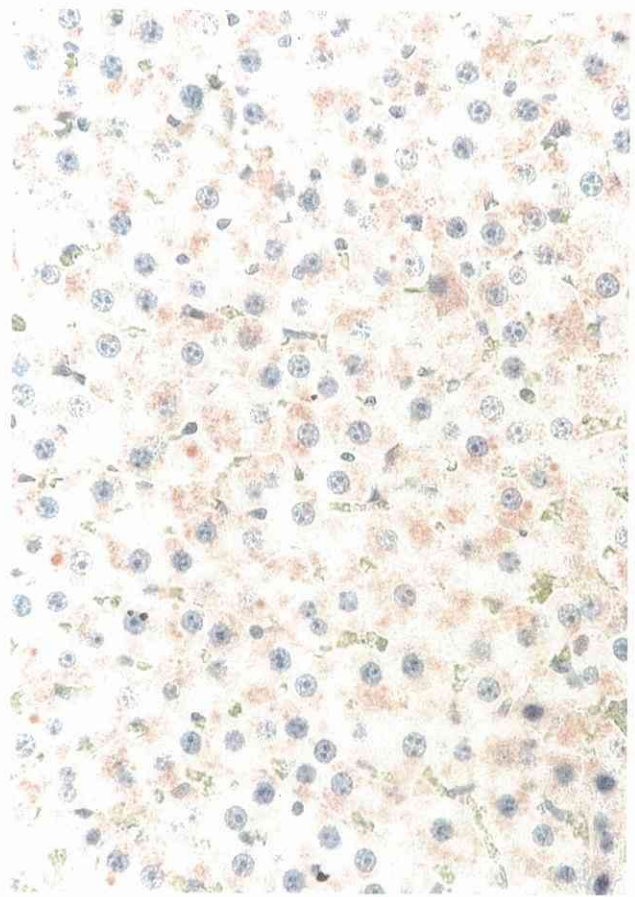


Figure 2 Effects of PFCA on the level of hepatic TG



Control



PFDA

Figure 3 Oil red O staining of the liver from PFDA-treated rats.

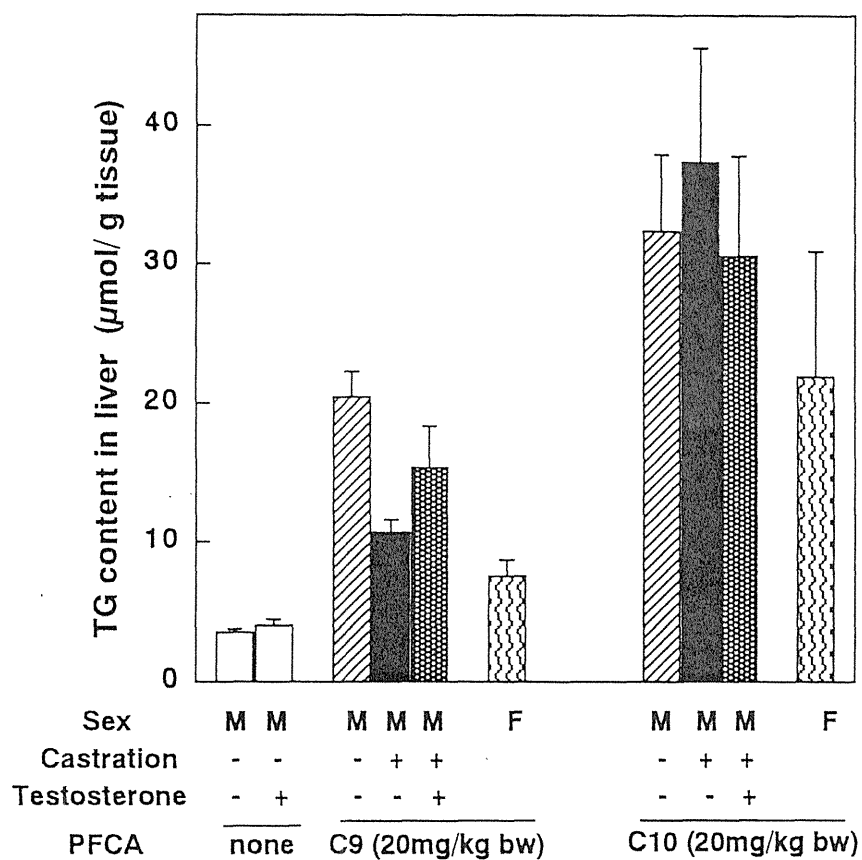


Figure 4 Effects of castration and testosterone on the level of triglyceride in the liver of male rats

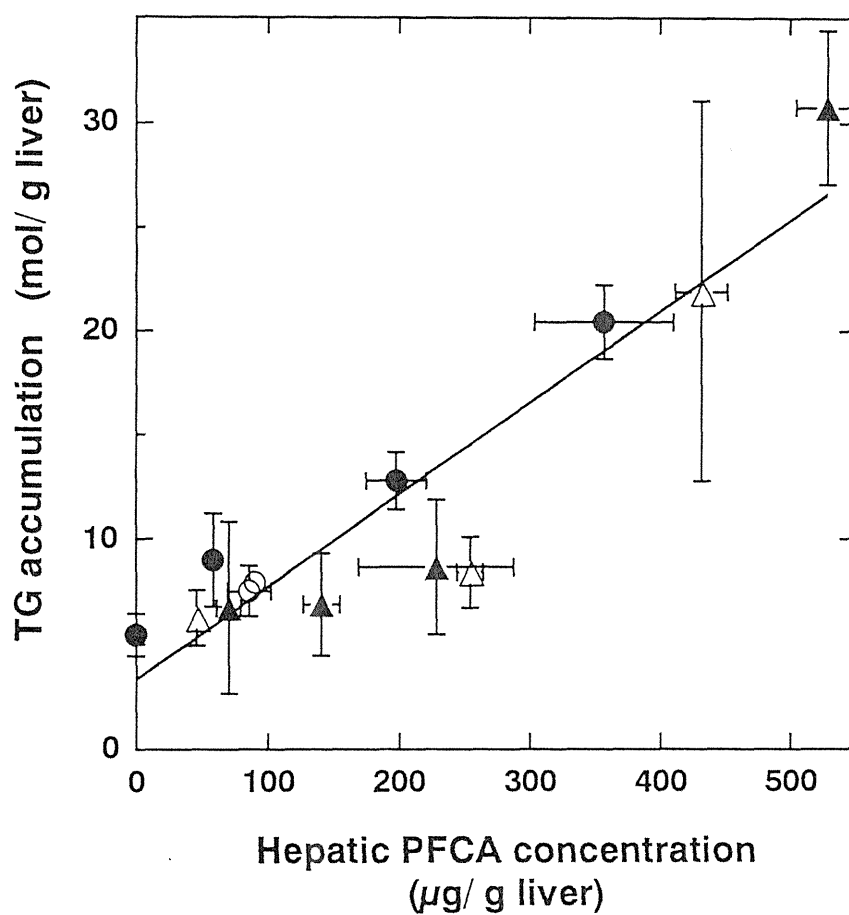


Figure 5 Relationship between PFCA concentration and the level of triglyceride in rat liver

Table 1 Effects of PFDA on fatty acid content of Liver Triglyceride

	% Composition		FA content/ g liver		FA content / whole liver	
	control	PFDA	control	PFDA	control	PFDA
Fatty acid	(% of total)		(μmol/ FA g liver)		(μmol FA/ liver)	
16:0	33.8± 1.7	32.0 ± 0.4	4.9 ± 1.1	29.6 ± 3.4	42.1 ±11.7	244.7 ± 50.2
16:1(n-7)	3.4 ± 0.9	3.2 ± 0.9	0.5 ± 0.2	2.8 ± 0.8	4.4 ± 1.9	25.4 ± 11.5
18:0	1.6 ± 0.2	1.6 ± 0.4	0.2 ± 0.1	1.4 ± 0.1	1.9 ± 0.5	12.3 ± 1.7
18:1(n-9)	22.3 ± 0.7	31.2 ± 1.9	3.3 ± 0.8	28.9 ± 4.5	27.9 ± 7.9	240.1 ± 61.7
18:2(n-6)	28.4 ± 1.9	27.5 ± 2.5	4.1 ± 0.8	25.2 ± 1.5	34.9 ± 8.6	206.8 ± 25.2
18:3(n-3)	1.4 ± 0.3	0.8 ± 0.1	0.2 ± 0.1	0.7 ± 0.1	1.7 ± 0.6	5.7 ± 1.2
20:3(n-6)	0.5 ± 0.1	0.5 ± 0.1	0.5 ± 0.1	0.4 ± 0.0	0.5 ± 0.1	3.7 ± 0.7
20:4(n-6)	1.5 ± 0.3	1.4 ± 0.2	0.2 ± 0.0	1.3 ± 0.1	1.9 ± 0.5	10.5 ± 1.7
20:5(n-3)	1.7 ± 0.3	0.3 ± 0.1	0.2 ± 0.1	0.3 ± 0.1	2.0 ± 0.6	2.2 ± 0.8
22:6(n-3)	3.7 ± 0.7	1.2 ± 0.1	0.5 ± 0.2	1.1 ± 0.1	4.5 ± 1.6	9.2 ± 1.7
Total	100.0 ± 0.0	100.0 ± 0.0	14.5 ± 3.0	92.2 ± 11.0	124.2 ± 32.9	763.7 ± 157.0

Male Wistar rats (6-week old) were intraperitoneally administered with perfluorodecanoic acid (C10) (20 mg/ kg body weight) once a day for 5 days before being killed.

Values represent mean ± SD for 4 rats.

